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Specified deleted sequences of tubercle bacilli and their uses in diagnostics and vaccines .

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The present invention pertains to the field of biology, more particularly the subject of the present invention is the identification of a nucleotide sequence which make it possible in particular to distinguish an infection resulting from *Mycobacterium tuberculosis* from an infection resulting from *Mycobacterium africanum*, *Mycobacterium canetti*, *Mycobacterium microti*, *Mycobacterium bovis*, *Mycobacterium bovis BCG*. The subject of the present invention is also a method for detecting the sequences in question by the products of expression of these sequences and the kits for carrying out these methods. Finally, the subject of the present invention is novel vaccines.

Despite more than a century of research since the discovery of *Mycobacterium tuberculosis*, the aetiological agent of tuberculosis, this disease remains one of the major causes of human mortality. *M. tuberculosis* is expected to kill 3 million people annually (Snider, 1989 Rev. Inf. Dis. S335) and the number of new people getting infected each year is rising and is estimated at 8.8 million. Although the majority of these are in developing countries, the disease is assuming renewed importance in the western countries due to the increasing number of homeless people, the impact of the AIDS epidemic, the changing global migration, and the travel patterns.

Early tuberculosis often goes unrecognized in an otherwise healthy individual. Classical initial methods of diagnosis include examination of a sputum smear under a microscope for acid-fast mycobacteria and an x-ray of the lungs. However, in a vast majority of cases the sputum smear examination is negative for Mycobacteria in the early stages of the disease, and lung changes may not be obvious on an x-ray until several months following infection. Another complicating factor is that acid-fast bacteria in a sputum smear may often be other species of mycobacteria. Antibiotics used for treating tuberculosis have considerable side effects, and must be taken as a combination of three or more drugs for a six to twelve month period. In addition, the possibility of inducing the appearance of drug resistant tuberculosis prevents therapy from being administered without solid evidence to support the diagnosis. Currently the only absolutely reliable method of diagnosis is based on culturing *M. tuberculosis* from the clinical specimen and identifying it morphologically and biochemically. This usually takes anywhere from three to six weeks, during which time a patient may become seriously ill and infect other individuals. Therefore, a rapid test capable of reliably detecting the presence of *M. tuberculosis* is vital for the early detection and treatment. Several molecular tests have been developed recently for the rapid detection and identification of *M. tuberculosis*, such as the Gen-Probe "Amplified *Mycobacterium tuberculosis* Direct Test"; this test amplifies *M. tuberculosis* 16S ribosomal RNA from

respiratory specimens and uses a chemiluminescent probe to detect the amplified product with a reported sensitivity of about 91%. The discovery of the IS6110 insertion element (Cave et al., Eisenach *et al.*, 1990 J. Infectious Diseases 161:977-981; Thierry *et al.* 1990 J. Clin. Microbiol. 28: 2668-2673) and the belief that this element may only be present in *Mycobacterium* complex (*M. tuberculosis*, *M. bovis*, *M. bovis*-BCG, *M. africanum* and *M. microti*) spawned a whole series of rapid diagnostic strategies (Brisson-Noel *et al.*, 1991 Lancet 338: 364-366; Clarridge *et al.* 1993, J. Clin. Microbiol. 31 :2049-2056 ; Cormican *et al.* 1992 J. Clin. Pathology 1992, 45 : 601-604 ; Cousins *et al.*, 1992 J. Clin. Microbiol. 30 : 255-258 ; Del Portillo *et al.* 1991 J. Clin. Microbiol. 29 : 2163-2168 ; Folgueira *et al.*, 1994 Neurology 44 :1336-1338 ; Forbes *et al.* 1993, J.Clin.Microbiol. 31 :1688-1694 ; Hermans *et al.* 1990 J. Clin. Microbiol. 28 :1204-1213 ; Kaltwasser *et al.* 1993 Mol. Cell. Probes 7 : 465-470 ; Kocagoz *et al.* 1993 J. Clin. Microbiol. 31 :1435-1438 ; Kolk *et al.* 1992 J.Clin.Microbiol. 30 : 2567-2575 ; Kox *et al.* 1994 J.Clin.Microbiol. 32 :672-678 ; Liu *et al.* 1994 Neurology 44 :1161-1164 ; Miller *et al.* 1994 J. Clin.Microbiol. 32 : 393-397 ; Reischl *et al.* 1994 Biotechniques 17 :844-845 ; Schluger *et al.* 1994 Chest 105 :1116-1121 ; Shawar *et al.* 1993 J. Clin. Microbiol. 31: 61-65; Wilson *et al.* 1993 J.Clin.Microbiol. 28: 2668-2673). These tests employ various techniques to extract DNA from the sputum. PCR is used to amplify IS6110 DNA sequences from the extracted DNA. The successful amplification of this DNA is considered to be an indicator of the presence of *M. tuberculosis* infection. U.S. Pat. Nos. 5,168,039 and 5,370,998 have been issued to Crawford *et al.* for the IS6110 based detection of tuberculosis. European patent EP 0,461,045 has been issued to Guesdon for the IS6110 based detection of tuberculosis.

Thus, these molecular assays used to detect *M. tuberculosis* depend on the IS6110 insertion sequence (about 10 copies) or the 16S ribosomal RNA (thousands of copies). However, these methods do not provide any information regarding the sub-type of the mycobacteria. Indeed several dozen species of Mycobacteria are known, and most are non-pathogenic for humans; tuberculosis is usually caused by infection due to *M. tuberculosis*, with a few cases being caused by *M. bovis*, *M. canettii*, and *M. africanum*. In order to choose an appropriate treatment and to conduct epidemiological investigations it is absolutely necessary to be able to rapidly and accurately identify isolates, i.e to distinguish the sub-type of mycobacteria of the *Mycobacterium* complex, originating from potential tuberculosis patients. That's the problem the present invention intends to solve.

The present invention provides an isolated or purified nucleic acid wherein said nucleic acid is selected from the group consisting of:

- a) SEQ ID N°1, named TbD1 (*M. tuberculosis* specific deletion 1);
- b) Nucleic acid having a sequence fully complementary to SEQ ID N°1.
- c) Nucleic acid fragment comprising at least 8, 15, 20, 25, 30, 50, 100, 250, 500, 750, 1000, 1500, 2000, 2500, 3000 consecutive nucleotides of SEQ ID N°1;
- d) Nucleic acid having at least 90% sequence identity after optimal alignment with a sequence defined in a) or b);
- e) Nucleic acid that hybridizes under stringent conditions with the nucleic acid defined in a) or b);

As used herein, the terms « isolated » and « purified » according to the invention refer to a level of purity that is achievable using current technology. The molecules of the invention do not need to be absolutely pure (i.e., contain absolutely no molecules of other cellular macromolecules), but should be sufficiently pure so that one of ordinary skill in the art would recognize that they are no longer present in the environment in which they were originally found (i.e., the cellular middle). Thus, a purified or isolated molecule according to the present invention is one that have been removed from at least one other macromolecule present in the natural environment in which it was found. More preferably, the molecules of the invention are essentially purified and/or isolated, which means that the composition in which they are present is almost completely, or even absolutely, free of other macromolecules found in the environment in which the molecules of the invention are originally found. Isolation and purification thus does not occur by addition or removal of salts, solvents, or elements of the periodic table, but must include the removal of at least some macromolecules. The nucleic acids encompassed by the invention are purified and/or isolated by any appropriate technique known to the ordinary artisan. Such techniques are widely known, commonly practiced, and well within the skill of the ordinary artisan. As used herein, the term "nucleic acid" refers to a polynucleotide sequence such as a single or double stranded DNA sequence, RNA sequence, cDNA sequence; such a polynucleotide sequence has been isolated, purified or synthesized and may be constituted with natural or non natural nucleotides. In a preferred embodiment the DNA molecule of the invention is a double stranded DNA molecule. As used herein, the terms "nucleic acid", "oligonucleotide", "polynucleotide" have the same meaning and are used indifferently.

By the term "*Mycobacterium* complex" as used herein, it is meant the complex of mycobacteria causing tuberculosis which are *Mycobacterium tuberculosis*, *Mycobacterium*

*bovis*, *Mycobacterium africanum*, *Mycobacterium microti*, *Mycobacterium canettii* and the vaccine strain *Mycobacterium bovis* BCG.

The present invention encompasses not only the entire sequence SEQ ID N°1, its complement, and its double-stranded form, but any fragment of this sequence, its  
5 complement, and its double-stranded form.

In embodiments, the fragment of SEQ ID N°1 comprises at least approximately 8 nucleotides. For example, the fragment can be between approximately 8 and 30 nucleotides and can be designed as a primer for polynucleotide synthesis. In another preferred  
10 embodiment, the fragment of SEQ ID N°1 comprises between approximately 1,500 and approximately 2,500 nucleotides, and more preferably 2153 nucleotides corresponding to SEQ ID N°4. As used herein, "nucleotides" is used in reference to the number of nucleotides on a single-stranded nucleic acid. However, the term also encompasses double-stranded molecules. Thus, a fragment comprising 2,153 nucleotides according to the invention is a single-stranded molecule comprising 2,153 nucleotides, and also a double stranded molecule  
15 comprising 2153 base pairs (bp).

In a preferred embodiment, the nucleic acid fragment of the invention is specifically deleted in the genome of *Mycobacterium tuberculosis*, excepted in *Mycobacterium tuberculosis* strain having the mutation CTG → CGG at codon 463 of gene KatG and/or having no or very few IS6110 sequences inserted in their genome and present in the genome  
20 of *Mycobacterium africanum*, *Mycobacterium canetti*, *Mycobacterium microti*, *Mycobacterium bovis*, *Mycobacterium bovis* BCG. By the term "few IS6110 sequences inserted in the genome", it is meant less than ten copies in the genome of *M. tuberculosis*, more preferably less than 5 copies, for example less than two copies.

The nucleic acid fragment of the invention is preferably selected from the group  
25 consisting of:

- a) SEQ ID N°4;
- b) Nucleic acid having a sequence fully complementary to SEQ ID N°4.
- c) Nucleic acid fragment comprising at least 20, 25, 30, 50, 100, 250, 500, 750, 1000, 1500, 2000, 2500, 3000 consecutive nucleotides of SEQ ID N°4;
- 30 d) Nucleic acid having at least 90% sequence identity after optimal alignment with a sequence defined in a) or b);
- e) Nucleic acid that hybridizes under stringent conditions with the nucleic acid defined in a) or b); In embodiments, the stringent conditions under which a sequence according to the invention is determined are conditions which are no less stringent than 5xSSPE,

2xDenhardt's solution, and 0.5% (w/v) sodium dodecyl sulfate at 65°C. More stringent conditions can be utilized by the ordinary artisan, and the proper conditions for a given assay can be easily and rapidly determined without undue or excessive experimentation. As an illustrative embodiment, the stringent hybridization conditions used in order to specifically  
5 detect a polynucleotide according to the present invention are advantageously the following: pre-hybridization and hybridization are performed at 65°C in a mixture containing:

- 5X SSPE (1X SSPE is 3 M NaCl, 30 mM tri-sodium citrate)
- 2X Denhardt's solution
- 0.5% (w/v) sodium dodecyl sulfate (SDS)
- 10 - 100 µg ml<sup>-1</sup> salmon sperm DNA.

The washings are performed as follows:

- two washings at laboratory temperature (approximately 21-25°C) for 10 min. in the presence of 2X SSPE and 0.1% SDS; and
- one washing at 65°C for 15 min. in the presence of 1X SSPE and 0.1% SDS.

15 The invention also encompasses the isolated or purified nucleic acid of the invention wherein said nucleic acid comprises at least a deletion of a nucleic acid fragment as defined above.

Polynucleotides of the invention can be characterized by the percentage of identity they show with the sequences disclosed herein. For example, polynucleotides having at least  
20 90% identity with the polynucleotides of the invention, particularly those sequences of the sequence listing, are encompassed by the invention. Preferably, the sequences show at least 90% identity with those of the sequence listing. More preferably, they show at least 92% identity, for example 95% or 99% identity. The skilled artisan can identify sequences according to the invention through the use of the sequence analysis software BLAST (see for  
25 example, Coffin et al., eds., "*Retroviruses*", Cold Spring Harbor Laboratory Press, pp. 723-755). Percent identity is calculated using the BLAST sequence analysis program suite, Version 2, available at the NCBI (NIH). All default parameters are used. BLAST (Basic Local Alignment Search Tool) is the heuristic search algorithm employed by the programs blastp, blastn, blastx, tblastn and tblastx, all of which are available through the BLAST  
30 analysis software suite at the NCBI. These programs ascribe significance to their findings using the statistical methods of Karlin and Altschul (1990, 1993) with a few enhancements. Using this publicly available sequence analysis program suite, the skilled artisan can easily identify polynucleotides according to the present invention.

It is well within the skill of the ordinary artisan to identify regions of the nucleic acid sequence of the invention, which would be useful as a probe, primer, or other experimental, diagnostic, or therapeutic aid. For example, the ordinary artisan could utilize any of the widely available sequence analysis programs to select regions (fragments) of these sequences that are useful for hybridization assays such as Southern blots, Northern blots, DNA binding assays, and/or *in vitro*, *in situ*, or *in vivo* hybridizations. Additionally, the ordinary artisan, with the sequences of the present invention, can utilize widely available sequence analysis programs to identify regions that can be used as probes and primers, as well as for design of anti-sense molecules. The only practical limitation on the fragment chosen by the ordinary artisan is the ability of the fragment to be useful for the purpose for which it is chosen. For example, if the ordinary artisan wished to choose a hybridization probe, he would know how to choose one of sufficient length, and of sufficient stability, to give meaningful results. The conditions chosen would be those typically used in hybridization assays developed for nucleic acid fragments of the approximate chosen length.

Thus, the present invention provides short oligonucleotides, such as those useful as probes and primers. In embodiments, the probe and/or primer comprises 8 to 30 consecutive nucleotides of the polynucleotide according to the invention or the polynucleotide complementary thereto. Advantageously, a fragment as defined herein has a length of at least 8 nucleotides, which is approximately the minimal length that has been determined to allow specific hybridization. Preferably the nucleic fragment has a length of at least 12 nucleotides and more preferably 20 consecutive nucleotides of any of SEQ ID NO:1 or SEQ ID NO:3. The sequence of the oligonucleotide can be any of the many possible sequences according to the invention. Preferably, the sequence is selected from the following group SEQ ID No. 13, SEQ ID No. 14, SEQ ID N°15, SEQ ID N°16, SEQ ID N°17, SEQ ID N°18. More precisely, the primers pairs SEQ ID N°13/SEQ ID N°14 and SEQ ID N°15/SEQ ID N°16 are specific for nucleic acid fragment SEQ ID N°4. The primers pair SEQ ID N°17/SEQ ID No. 18 is specific for nucleic acid sequence SEQ ID N°1 and are flanking the nucleic acid fragment of SEQ ID N°4.

Thus, the polynucleotides of SEQ ID N°1 and SEQ ID N°4, and their fragments, can be used to select nucleotide primers, notably for an amplification reaction, such as the amplification reactions further described.

PCR is described in US Patent No. 4,683,202, which is incorporated in its entirety herein. The amplified fragments may be identified by agarose or polyacrylamide gel electrophoresis, by a capillary electrophoresis, or alternatively by a chromatography



technique (gel filtration, hydrophobic chromatography, or ion exchange chromatography). The specificity of the amplification can be ensured by a molecular hybridization using as nucleic probes the polynucleotides of SEQ ID N°1 or SEQ ID N°4, and their fragments, oligonucleotides that are complementary to these polynucleotides or fragments thereof, or  
 5 their amplification products themselves, or even by DNA sequencing.

The following other techniques related to nucleic acid amplification may also be used and are generally preferred to the PCR technique. The Strand Displacement Amplification (SDA) technique is an isothermal amplification technique based on the ability of a restriction enzyme to cleave one of the strands at a recognition site (which is under a  
 10 hemiphosphorothioate form) and on the property of a DNA polymerase to initiate the synthesis of a new strand from the 3'OH end generated by the restriction enzyme and on the property of this DNA polymerase to displace the previously synthesized strand being localized downstream. The SDA amplification technique is more easily performed than PCR (a single thermostatted water bath device is necessary), and is faster than the other  
 15 amplification methods. Thus, the present invention also comprises using the nucleic acid fragments according to the invention (primers) in a method of DNA or RNA amplification according to the SDA technique.

When the target polynucleotide to be detected is a RNA, for example a mRNA, a reverse transcriptase enzyme will be used before the amplification reaction in order to obtain  
 20 a cDNA from the RNA contained in the biological sample. The generated cDNA is subsequently used as the nucleic acid target for the primers or the probes used in an amplification process or a detection process according to the present invention.

The non-labeled polynucleotides or oligonucleotides of the invention can be directly used as probes. Nevertheless, the polynucleotides or oligonucleotides are generally labeled  
 25 with a radioactive element ( $^{32}\text{P}$ ,  $^{35}\text{S}$ ,  $^3\text{H}$ ,  $^{125}\text{I}$ ) or by a non-isotopic molecule (for example, biotin, acetylaminofluorene, digoxigenin, 5-bromodesoxyuridine, fluorescein) in order to generate probes that are useful for numerous applications. Examples of non-radioactive labeling of nucleic acid fragments are described in French patent N° FR 78 10975 and by Urdea *et al.* (1988, *Nucleic Acids Research* 11:4937-4957) or Sanchez-Pescador *et al.* (1988,  
 30 *J. Clin. Microbiol.* 26(10):1934-1938), the disclosures of which are hereby incorporated in their entirety. Other labeling techniques can also be used, such as those described in French patents FR 2 422 956 and FR 2 518 755. The hybridization step may be performed in different ways. See, for example, Matthews *et al.*, 1988, *Anal. Biochem.* 169:1-25. A general method comprises immobilizing the nucleic acid that has been extracted from the biological

sample on a substrate (for example, nitrocellulose, nylon, polystyrene) and then incubating, in defined conditions, the target nucleic acid with the probe. Subsequent to the hybridization step, the excess amount of the specific probe is discarded and the hybrid molecules formed are detected by an appropriate method (radioactivity, fluorescence or enzyme activity measurement, etc.).

Amplified nucleotide fragments are useful, among other things, as probes used in hybridization reactions in order to detect the presence of one polynucleotide according to the present invention or in order to detect mutations. The primers may also be used as oligonucleotide probes to specifically detect a polynucleotide according to the invention.

The oligonucleotide probes according to the present invention may also be used in a detection device comprising a matrix library of probes immobilized on a substrate, the sequence of each probe of a given length being localized in a shift of one or several bases, one from the other, each probe of the matrix library thus being complementary to a distinct sequence of the target nucleic acid. Optionally, the substrate of the matrix may be a material able to act as an electron donor, the detection of the matrix positions in which an hybridization has occurred being subsequently determined by an electronic device. Such matrix libraries of probes and methods of specific detection of a target nucleic acid is described in the European patent application N° EP-0 713 016 (Affymax technologies) and also in the US patent N° US-5,202,231 (Drmanac). Since almost the whole length of a mycobacterial chromosome is covered by BAC-based genomic DNA library (i.e. 97% of the *M. tuberculosis* chromosome is covered by the BAC library I-1945), these DNA libraries will play an important role in a plurality of post-genomic applications, such as in mycobacterial gene expression studies where the canonical set of BACs could be used as a matrix for hybridization studies. Thus it is also in the scope of the invention to provide a nucleic acid chips, more precisely a DNA chips or a protein chips that respectively comprises a nucleic acid or a polypeptide of the invention.

The present invention is also providing a vector comprising the isolated DNA molecule of the invention. A "vector" is a replicon in which another polynucleotide segment is attached, so as to bring the replication and/or expression to the attached segment. A vector can have one or more restriction endonuclease recognition sites at which the DNA sequences can be cut in a determinable fashion without loss of an essential biological function of the vector, and into which a DNA fragment can be spliced in order to bring about its replication and cloning. Vectors can further provide primer sites (e.g. for PCR), transcriptional and/or translational initiation and/or regulation sites, recombinational signals,

replicons, selectable markers, etc. Beside the use of homologous recombination or restriction enzymes to insert a desired DNA fragment into the vector, UDG cloning of PCR fragments (US Pat. No. 5,334,575), T:A cloning, and the like can also be applied. The cloning vector can further contain a selectable marker suitable for use in the identification of cells transformed with the cloning vector.

The vector can be any useful vector known to the ordinary artisan, including, but not limited to, a cloning vector, an insertion vector, or an expression vector. Examples of vectors include plasmids, phages, cosmids, phagemid, yeast artificial chromosome (YAC), bacterial artificial chromosome (BAC), human artificial chromosome (HAC), viral vector, such as adenoviral vector, retroviral vector, and other DNA sequences which are able to replicate or to be replicated *in vitro* or in a host cell, or to convey a desired DNA segment to a desired location within a host cell. According to a preferred embodiment of the invention, the recombinant vector is a BAC pBeloBAC11 in which the genomic region of *Mycobacterium bovis*-BCG 1173P3 that spans the region corresponding to the locus 1,760,753 bp to 1,830,364 bp in the genome of *M. tuberculosis* H37Rv has been inserted into the HindIII restriction site; this recombinant vector is named X229. In this region, the inventors have demonstrated the deletion of a 2153 bp fragment in the vast majority of *M. tuberculosis* strain. That's the reason why the inventors named this region TbD1 ("*M. tuberculosis* specific deletion 1"). This 2153 bp region is flanked by the sequence GGC CTG GTC AAA CGC GGC TGG ATG CTG and AGA TCC GTC TTT GAC ACG ATC GAC.G. External primers hybridizing with such sequences or the complementary sequences thereof can be used for the amplification of TbD1 region to check for the presence or the absence of the deletion of the TbD1 region. The inventors design for example the following primers:

5'- CTA CCT CAT CTT CCG GTC CA-3' (SEQ ID N° 17)

5'- CAT AGA TCC CGG ACA TGG TG-3' (SEQ ID N° 18)

In order to get a specific 500 pb probe for hybridization experiments, a PCR amplification of an internal fragment may be realized by using the plasmid X229 as a matrix. The amplification of a fragment of approximatively 500 bp internal to the TbD1 region can be performed by using the following primers:

5'- CGT TCA ACC CCA AAC AGG TA-3' (SEQ ID N° 13)

5'- AAT CGA ACT CGT GGA ACA CC-3' (SEQ ID N° 14)

The amplification of a fragment of approximatively 2,000 bp internal to the TbD1 region can be performed by using the following primers:

5'- ATT CAG CGT CTA TCG GTT GC-3' (SEQ ID N° 15)

5'- AGC AGC TCG GGA TAT CGT AG-3' (SEQ ID N° 16)

The PCR conditions are the following: denaturation 95°C 1 min, then 35 cycles of amplification [95°C during 30 seconds, 58°C during 1 min] , then elongation 72°C during 4 min.

5        Thus, this invention also concerns a recombinant cell host which contains a polynucleotide or recombinant vector according to the invention. The cell host can be transformed or transfected with a polynucleotide or recombinant vector to provide transient, stable, or controlled expression of the desired polynucleotide. For example, the polynucleotide of interest can be subcloned into an expression plasmid at a cloning site  
10       downstream from a promoter in the plasmid and the plasmid can be introduced into a host cell where expression can occur. The recombinant host cell can be any suitable host known to the skilled artisan, such as a eukaryotic cell or a microorganism. For example, the host can be a cell selected from the group consisting of *Escherichia coli*, *Bacillus subtilis*, insect cells, and yeasts. According to a preferred embodiment of the invention, the recombinant cell host  
15       is a commercially available *Escherichia coli* DH10B (Gibco) containing the BAC named X229 previously described. This *Escherichia coli* DH10B (Gibco) containing the BAC named X229 has been deposited with the Collection Nationale de Cultures de Microorganismes (CNCM), Institut Pasteur, Paris, France, on February 18<sup>th</sup>, 2002 under number CNCM I-2799.

20       Another aspect of the invention is the isolated or purified polypeptides encoded by a polynucleotide of the invention. The purified polypeptide comprises an amino acid sequence that is encoded by SEQ ID N° 6, N° 8, N° 10, N° 12, and their fragments thereof. For example, the purified polypeptide of the invention can comprise the amino acid sequence of SEQ ID N° 6, which is the amino acid sequence of the mmpL6 protein or the amino acid  
25       sequence of SEQ ID N° 8 a truncated form of mmpL6. The purified polypeptide of the invention can comprise the amino acid sequence of SEQ ID NO:10, which is the amino acid sequence of the mmpS6 protein or the amino acid sequence of SEQ ID N° 12 a truncated form of mmpS6.

30       It is now easy to produce proteins in large amounts by genetic engineering techniques through the use of expression vectors, such as plasmids, phages, and phagemids. The polypeptide of the present invention can be produced by insertion of the appropriate polynucleotide into an appropriate expression vector at the appropriate position within the vector. Such manipulation of polynucleotides is well known and widely practiced by the ordinary artisan. The polypeptide can be produced from these recombinant vectors either *in*

*vitro* or *in vivo*. All the isolated or purified nucleic acid encoding by the polypeptide of the invention are in the scope of the invention. The polypeptide of the invention is a polypeptide encoded by a polynucleotide which hybridizes to any of SEQ ID N°1 or N°4 under stringent conditions, as defined herein.

5 More preferably, said isolated or purified nucleic acid according the invention is selected among:

- SEQ ID N° 5 encoding the polypeptide of SEQ ID N°6;
- SEQ ID N° 7 encoding the polypeptide of SEQ ID N°8;
- SEQ ID N° 9 encoding the polypeptide of SEQ ID N°10;
- 10 - SEQ ID N° 11 encoding the polypeptide of SEQ ID N°12.

The present invention also provides a method for the discriminatory detection and identification of:

- *Mycobacterium tuberculosis* excepted *Mycobacterium tuberculosis* strain having the mutation CTG  $\rightarrow$  CGG at codon 463 of gene KatG and/or excepted *Mycobacterium tuberculosis* strain having no or very few IS6110 sequences inserted in their genome; versus,
  - 15 - *Mycobacterium africanum*, *Mycobacterium canettii*, *Mycobacterium microti*, *Mycobacterium bovis*, *Mycobacterium bovis BCG* in a biological sample,
- comprising the following steps:

- a) isolation of the DNA from the biological sample to be analyzed or
- 20 production of a cDNA from the RNA of the biological sample,
- b) detection of the nucleic acid sequences of the mycobacterium present in said biological sample,
- c) analysis for the presence or the absence of a nucleic acid fragment as previously described.

25 By a biological sample according to the present invention, it is notably intended a biological fluid, such as sputum, saliva, plasma, blood, urine or sperm, or a tissue, such as a biopsy.

Analysis of the desired sequences may, for example, be carried out by agarose gel electrophoresis. If the presence of a DNA fragment migrating to the expected site is observed, it can be concluded that the analyzed sample contained mycobacterial DNA. This

30 analysis can also be carried out by the molecular hybridization technique using a nucleic probe. This probe will be advantageously labeled with a nonradioactive (cold probe) or radioactive element. Advantageously, the detection of the mycobacterial DNA sequences will be carried out using nucleotide sequences complementary to said DNA sequences. By way of example, they may include labeled or nonlabeled nucleotide probes; they may also

include primers for amplification. The amplification technique used may be PCR but also other alternative techniques such as the SDA (Strand Displacement Amplification) technique, the TAS technique (Transcription-based Amplification System), the NASBA (Nucleic Acid Sequence Based Amplification) technique or the TMA (Transcription Mediated Amplification) technique.

The primers in accordance with the invention have a nucleotide sequence chosen from the group comprising SEQ ID No. 13, SEQ ID No. 1, SEQ ID N°15, SEQ ID N°16, SEQ ID N°17, SEQ ID N°18. The pairs SEQ ID N°13/SEQ ID N°14 and SEQ ID N°15/SEQ ID N°16 specific for nucleic acid fragment SEQ ID N°4, and the pair SEQ ID N°17/SEQ ID N°18 specific for nucleic acid of the invention.

In a variant, the subject of the invention is also a method for the discriminatory detection and identification of:

- *Mycobacterium tuberculosis* excepted *Mycobacterium tuberculosis* strain having the mutation CTG -> CGG at codon 463 of gene katG and/or excepted *Mycobacterium tuberculosis* strain having no or very few IS6110 sequences inserted in their genome; versus,
  - *Mycobacterium africanum*, *Mycobacterium canettii*, *Mycobacterium microti*, *Mycobacterium bovis*, *Mycobacterium bovis BCG* in a biological sample,
- comprising the following steps:

- a) bringing the biological sample to be analyzed into contact with at least one pair of primers as defined in claim 11 or 12, the DNA contained in the sample having been, where appropriate, made accessible to the hybridization beforehand,
- b) amplification of the DNA of the mycobacterium,
- c) visualization of the amplification of the DNA fragments.

The amplified fragments may be identified by agarose or polyacrylamide gel electrophoresis by capillary electrophoresis or by a chromatographic technique (gel filtration, hydrophobic chromatography or ion-exchange chromatography). The specification of the amplification may be controlled by molecular hybridization using probes, plasmids containing these sequences or their product of amplification. The amplified nucleotide fragments may be used as reagent in hybridization reactions in order to detect the presence, in a biological sample, of a target nucleic acid having sequences complementary to those of said amplified nucleotide fragments. These probes and amplicons may be labeled or otherwise with radioactive elements or with nonradioactive molecules such as enzymes or fluorescent elements.

The subject of the present invention is also a kit for the discriminatory detection and identification of:

- *Mycobacterium tuberculosis* excepted *Mycobacterium tuberculosis* strain having the mutation CTG → CGG at codon 463 of gene katG and/or excepted *Mycobacterium tuberculosis* strain having no or very few IS6110 sequences inserted in their genome; versus,
  - *Mycobacterium africanum*, *Mycobacterium canettii*, *Mycobacterium microti*, *Mycobacterium bovis*, *Mycobacterium bovis BCG* in a biological sample,
- in a biological sample comprising the following elements:

- a) at least one pair of primers as defined previously,
- b) the reagents necessary to carry out a DNA amplification reaction,
- c) optionally, the necessary components which make it possible to verify or compare the sequence and/or the size of the amplified fragment.

Indeed, in the context of the present invention, depending on the pair of primers used, it is possible to obtain very different results. Thus, the use of primers which are internal to the deletion, such as for example SEQ ID N°13, SEQ ID N°14, SEQ ID N°15, SEQ ID N°16, is such that no amplification product is detectable in *M. tuberculosis* and that amplification product is detectable in *Mycobacterium africanum*, *Mycobacterium canettii*, *Mycobacterium microti*, *Mycobacterium bovis*, *Mycobacterium bovis BCG*, *Mycobacterium tuberculosis* having the mutation CTG → CGG at codon 463 of gene katG and/or having no or very few IS6110 sequences inserted in their genome. However, the use of primers external to the region of deletion does not necessarily give the same result, as regards for example the size of the amplified fragment, depending on the size of the deleted region in *M. tuberculosis*. Thus, the use of the pair of primers external to the deletion such as SEQ ID N° 17 and SEQ ID N° 18 is likely to give rise to an amplicon in *Mycobacterium africanum*, *Mycobacterium canettii*, *Mycobacterium microti*, *Mycobacterium bovis*, *Mycobacterium bovis BCG*, *Mycobacterium tuberculosis* having the mutation CTG → CGG at codon 463 of gene katG and/or having no or very few IS6110 sequences inserted in their genome, of about 2100 bp whereas the use of the pair of primers external to the deletion will give rise in *M. tuberculosis* to an amplicon of about few bp.

More generally, the invention pertains to the use of at least one pair of primers as defined previously for the amplification of a DNA sequence from *Mycobacterium tuberculosis* or *Mycobacterium africanum*, *Mycobacterium canettii*, *Mycobacterium microti*, *Mycobacterium bovis*, *Mycobacterium bovis BCG*, *Mycobacterium Tuberculosis* having the

mutation CTG → CGG at codon 463 of gene katG and/or having no or very few IS6110 sequences inserted in their genome.

The subject of the invention is also the product of expression of all or part of the nucleic acid fragment as defined previously and deleted from the genome of *Mycobacterium tuberculosis* and present in *Mycobacterium africanum*, *Mycobacterium canettii*,  
 5 *Mycobacterium microti*, *Mycobacterium bovis*, *Mycobacterium bovis BCG*, *Mycobacterium Tuberculosis* having the mutation CTG → CGG at codon 463 of gene katG and/or having no or very few IS6110 sequences inserted in their genome; or conversely. The expression

“product of expression” is understood to mean any protein, polypeptide or polypeptide  
 10 fragment resulting from the expression of all or part of the above-mentioned nucleotide sequences. Among those product of expression, one can cite the membrane proteins mmpL6, mmpS6, and their truncated or rearranged form due to the deletion of the fragment of the invention.

Indeed, the subject of the present invention is also a method for the discriminatory  
 15 detection *in vitro* of antibodies directed against *Mycobacterium tuberculosis* or *Mycobacterium africanum*, *Mycobacterium canettii*, *Mycobacterium microti*, *Mycobacterium bovis*, *Mycobacterium bovis BCG*, *Mycobacterium tuberculosis* having the mutation CTG → CGG at codon 463 of gene KatG and/or having no or very few IS6110 sequences inserted in their genome, in a biological sample, comprising the following steps:

- 20 a) bringing the biological sample into contact with at least one product as previously defined ,
- b) detecting the antigen-antibody complex formed.

The subject of the present invention is also a method for the discriminatory detection of a vaccination with *Mycobacterium bovis BCG* or an infection by  
 25 *Mycobacterium tuberculosis*, excepted *Mycobacterium tuberculosis* strain having the mutation CTG → CGG at codon 463 of gene katG and/or excepted *Mycobacterium tuberculosis* strain having no or very few IS6110 sequences inserted in their genome in a mammal, comprising the following steps:

- a) preparation of a biological sample containing cells, more particularly cells of  
 30 the immune system of said mammal and more particularly T cells,
- b) incubation of the biological sample of step a) with at least one product as previously defined ,



c) detection of a cellular reaction indicating prior sensitization of the mammal to said product, in particular cell proliferation and/or synthesis of proteins such as gamma-interferon. Cell proliferation may be measured, for example, by incorporating  $^3\text{H}$ -Thymidine.

The invention also relates to a kit for the *in vitro* diagnosis of an *Mycobacterium tuberculosis* infection, excepted infection with *Mycobacterium tuberculosis* strain having the mutation CTG  $\rightarrow$  CGG at codon 463 of gene katG and/or having no or very few IS6110 sequences inserted in their genome, in a mammal optionally vaccinated beforehand with *M. bovis* BCG comprising:

- a) a product as previously defined ,
- 10 b) where appropriate, the reagents for the constitution of the medium suitable for the immunological reaction,
- c) the reagents allowing the detection of the antigen-antibody complexes produced by the immunological reaction,
- d) where appropriate, a reference biological sample (negative control) free of
- 15 antibodies recognized by said product,
- e) where appropriate, a reference biological sample (positive control) containing a predetermined quantity of antibodies recognized by said product.

The reagents allowing the detection of the antigen-antibody complexes may carry a marker or may be capable of being recognized in turn by a labeled reagent, more particularly in the

20 case where the antibody used is not labeled.

The subject of the invention is also mono- or polyclonal antibodies, their chimeric fragments or antibodies, capable of specifically recognizing a product of expression in accordance with the present invention.

The present invention therefore also relates to a method for the discriminatory

25 detection of the presence of an antigen of *Mycobacterium tuberculosis* or *Mycobacterium africanum*, *Mycobacterium canettii*, *Mycobacterium microti*, *Mycobacterium bovis*, *Mycobacterium bovis*-BCG, *Mycobacterium Tuberculosis* having the mutation CTG  $\rightarrow$  CGG at codon 463 of gene katG and/or having no or very few IS6110-sequences inserted in their genome in a biological sample comprising the following steps:

- 30 a) bringing the biological sample into contact with an antibody of the invention,
- b) detecting the antigen-antibody complex formed.

The invention also relates to the kit for the discriminatory detection of the presence of an antigen of *Mycobacterium tuberculosis* or *Mycobacterium africanum*, *Mycobacterium canettii*, *Mycobacterium microti*, *Mycobacterium bovis*, *Mycobacterium bovis* BCG,

*Mycobacterium tuberculosis* having the mutation CTG → CGG at codon 463 of gene katG and/or having no or very few IS6110 sequences inserted in their genome, in a biological sample comprising the following steps:

- a) an antibody as previously claimed ,
- 5        b) the reagents for constituting the medium suitable for the immunological reaction,
- c) the reagents allowing the detection of the antigen-antibody complexes produced by the immunological reaction.

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The above-mentioned reagents are well known to a person skilled in the art who will have no difficulty adapting them to the context of the present invention.

- 10        The subject of the invention is also an immunological composition, characterized in that it comprises at least one product of expression in accordance with the invention.

Advantageously, the immunological composition in accordance with the invention enters into the composition of a vaccine when it is provided in combination with a pharmaceutically acceptable vehicle and optionally with one or more immunity adjuvant(s) such as alum or a representative of the family of muramylpeptides or incomplete Freund's

- 15        adjuvant.

The invention also relates to a vaccine comprising at least one product of expression in accordance with the invention in combination with a pharmaceutically compatible vehicle and, where appropriate, one or more appropriate immunity adjuvant(s).

- 20        The invention also provide an in vitro method for the detection and identification of *Mycobacterium tuberculosis* excepted *Mycobacterium tuberculosis* strain having the mutation CTG → CGG at codon 463 of gene katG and/or excepted *Mycobacterium tuberculosis* strain having no or very few IS6110 sequences inserted in their genome in a biological sample,

- 25        comprising the following steps:

- a)        isolation of the DNA from the biological sample to be analyzed or production of a cDNA from the RNA of the biological sample,
- b)        detection of the nucleic acid sequences of the mycobacterium present in said biological sample,
- 30        c) analysis for the presence or the absence of a nucleic acid fragment of the invention.

In another embodiment, the invention provides an *in vitro* method for the detection and identification of *Mycobacterium tuberculosis* excepted *Mycobacterium tuberculosis* strain having the mutation CTG → CGG at codon 463 of gene katG and/or excepted

*Mycobacterium tuberculosis* strain having no or very few IS6110 sequences inserted in their genome in a biological sample, comprising the following steps:

- a) bringing the biological sample to be analyzed into contact with at least one pair of primers selected among nucleic acid fragments of the invention, and more preferably selected among the primers chosen from the group comprising SEQ ID N°13, SEQ ID N°14, SEQ ID N°15, SEQ ID N°16, SEQ ID N°17, SEQ ID N°18, the DNA contained in the sample having been, where appropriate, made accessible to the hybridization beforehand,
- b) amplification of the DNA of the mycobacterium,
- c) visualization of the amplification of the DNA fragments.

The invention also provides a kit for the detection and identification of *Mycobacterium tuberculosis* excepted *Mycobacterium tuberculosis* strain having the mutation CTG → CGG at codon 463 of gene katG and/or excepted *Mycobacterium tuberculosis* strain having no or very few IS6110 sequences inserted in their genome in a biological sample, comprising the following elements:

- a) at least one pair of primers selected among nucleic acid fragments of the invention, and more preferably selected among the primers chosen from the group comprising SEQ ID N°13, SEQ ID N°14, SEQ ID N°15, SEQ ID N°16, SEQ ID N°17, SEQ ID N°18,
- b) the reagents necessary to carry out a DNA amplification reaction,
- c) optionally, the necessary components which make it possible to verify or compare the sequence and/or the size of the amplified fragment.

The invention also relates to a method for the detection *in vitro* of antibodies directed against *Mycobacterium tuberculosis* excepted *Mycobacterium Tuberculosis* having the mutation CTG → CGG at codon 463 of gene KatG and/or having no or very few IS6110 sequences inserted in their genome, in a biological sample, comprising the following steps:

- a) bringing the biological sample into contact with at least one product as defined previously,
- b) detecting the antigen-antibody complex formed.

It is also a goal of the invention to use the TbD1 deletion as a as a genetic marker for the differentiation of *Mycobacterium* strain of *Mycobacterium tuberculosis* complex.

It is also a goal of the invention to use mmpL6<sup>551</sup> polymorphism as a genetic marker (see SEQ ID N°20) for the differentiation of *Mycobacterium* strain of *Mycobacterium tuberculosis* complex.

The use of such genetic marker(s) in association with at least one genetic markers selected among RD1, RD2, RD3, RD4, RD5, RD6, RD7, RD8, RD9, RD10, Rd11, RD13, RD14, RvD1, RvD2, RvD3, RvD4, RvD5, katG<sup>463</sup>, gyrA<sup>95</sup>, oxyR<sup>285</sup>, pncA<sup>57</sup> allows the differentiation of Mycobacterium strain of Mycobacterium Tuberculosis complex (see example 4).

The present invention provides an *in vitro* method for the detection and identification of *Mycobacteria* from the *Mycobacterium Tuberculosis* complex in a biological sample, comprising the following steps:

- a) analysis for the presence or the absence of a nucleic acid fragment of the sequence TbD1, and
- b) analysis of at least one additional genetic marker selected among RD1, RD2, RD3, RD4, RD5, RD6, RD7, RD8, RD9, RD10, Rd11, RD13, RD14, RvD1, RvD2, RvD3, RvD4, RvD5, katG<sup>463</sup>, gyrA<sup>95</sup>, oxyR<sup>285</sup>, pncA<sup>57</sup>.

In a preferred embodiment, two additional markers are used, preferably RD4 and RD9. The analysis is performed by a technique selected among sequence hybridization, nucleic acid amplification, antigen-antibody complex.

It is also a goal of the present invention to provide a kit for the detection and identification of *Mycobacteria* from the *Mycobacterium Tuberculosis* complex in a biological sample comprising the following elements:

- a) at least one pair of primers selected among nucleic acid fragments of the invention, and more preferably selected among the primers chosen from the group comprising SEQ ID N°13, SEQ ID N°14, SEQ ID N°15, SEQ ID N°16, SEQ ID N°17, SEQ ID N°18,
- b) at least one pair of primers specific of the genetic markers selected among RD1, RD2, RD3, RD4, RD5, RD6, RD7, RD8, RD9, RD10, Rd11, RD13, RD14, RvD1, RvD2, RvD3, RvD4, RvD5, katG<sup>463</sup>, gyrA<sup>95</sup>, oxyR<sup>285</sup>, pncA<sup>57</sup>.
- c) the reagents necessary to carry out a DNA amplification reaction,
- d) optionally, the necessary components which make it possible to verify or compare the sequence and/or the size of the amplified fragment.

In a preferred embodiment, the kit comprises the following elements:

- a) at least one pair of primers selected among nucleic acid fragments of the invention, and more preferably selected among the primers chosen from the group comprising SEQ ID N°13, SEQ ID N°14, SEQ ID N°15, SEQ ID N°16, SEQ ID N°17, SEQ ID N°18,

- b) one pair of primers specific of the genetic markers RD4,
- c) one pair of primers specific of the genetic markers RD9,
- d) the reagents necessary to carry out a DNA amplification reaction,
- e) optionally, the necessary components which make it possible to verify or compare the sequence and/or the size of the amplified fragment.

The figures and examples presented below are provided as further guide to the practitioner of ordinary skill in the art and are not to be construed as limiting the invention in anyway.

## FIGURES

**Figure 1 :** Amplicons obtained from strains that have the indicated genomic region present or deleted. Sizes of amplicons in each group are uniform. Numbers correspond to strain designation used in Kremer et al. (1999, J. Clin Microbiol. 37: 2607-2618) (Ref. 8) and Supply et al (2001, J. Clin. Microbiol. 39: 3563-3571) (ref.9).

**Figure. 2:** Sequences in the TbD1 region obtained from strains of various geographic regions.

\* refers to groups based on *katG*<sup>c463</sup>/*gyrA*<sup>c95</sup> sequence polymorphism defined by Sreevatsan and colleagues (Ref. 2). Numbers correspond to strain designation used in Kremer et al. (1999, J. Clin Microbiol. 37: 2607-2618) (Ref. 8) and Supply et al (2001, J. Clin. Microbiol. 39: 3563-3571) (ref.9).

**Figure 3:** Spoligotypes of selected *M. tuberculosis* and *M. bovis* strains. Numbers correspond to strain designation used in Kremer et al. (1999, J. Clin Microbiol. 37: 2607-2618) (Ref. 8) and Supply et al (2001, J. Clin. Microbiol. 39: 3563-3571) (ref.9).

**Figure 4:** Scheme of the proposed evolutionary pathway of the tubercle bacilli illustrating successive loss of DNA in certain lineages (grey boxes) The scheme is based on presence or absence of conserved deleted regions and on sequence polymorphisms in five selected genes.

Note that the distances between certain branches may not correspond to actual phylogenetic differences calculated by other methods.

Blue arrows indicate that strains are characterized by *katG*<sup>c463</sup> CTG (Leu), *gyrA*<sup>c95</sup> ACC (Thr), typical for group 1 organisms. Green arrows indicate that strains belong to group 2 characterized by *katG*<sup>c463</sup> CGG (Arg), *gyrA*<sup>c95</sup> ACC (Thr). The red arrow indicates that strains belong to group 3, characterized by *katG*<sup>c463</sup> CGG (Arg), *gyrA*<sup>c95</sup> AGC (Ser), as defined by Sreevatsan and colleagues (Sreevastan et al., 1997 Proc. Natl. Acad.Sci USA 151: 9869-9874) (Ref. 2).

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## EXAMPLES

### 1. MATERIAL AND METHODS:

**1.1. Bacterial Strains:** The 100 *M. tuberculosis* complex strains comprised 46 *M. tuberculosis* strains isolated in 30 countries, 14 *M. africanum* strains, 28 *M. bovis* strains originating in 5 countries, 2 *M. bovis* BCG vaccine strains (Pasteur and Japan), 5 *M. microti* strains, and 5 *M. canettii* strains. The strains were isolated from human and animal sources and were selected to represent a wide diversity including 60 strains that have been used in a multi-center study (8). The *M. africanum* strains were retrieved from the collection of the Wadsworth Center, New York State Department of Health, Albany, New York, whereas the majority of the *M. bovis* isolates came from the collection of the University of Zaragoza, Spain. Four *M. canettii* strains are from the culture collection of the Institut Pasteur, Paris, France. The strains have been extensively characterized by reference typing methods, i.e. IS6110 restriction fragment length polymorphism (RFLP) typing and spoligotyping. *M. tuberculosis* H37Rv, *M. tuberculosis* H37Ra, *M. tuberculosis* CDC1551, *M. bovis* AF2122/97, *M. microti* OV254, and *M. canettii* CIPT 140010059 were included as reference strains. DNA was prepared as previously described (10).

### 1.2. Genome comparisons and primer design

For preliminary genome comparisons between *M. tuberculosis* and *M. bovis* websites <http://genolist.pasteur.fr/TubercuList/> and [http://www.sanger.ac.uk/Projects/M\\_bovis/](http://www.sanger.ac.uk/Projects/M_bovis/) as well as inhouse databases were used. For primer design, sequences inside or flanking RD and RvD regions were obtained from the same websites. Primers were designed using the

primer 3 website [http://www-genome.wi.mit.edu/cgi-bin/primer/primer3\\_www.cgi](http://www-genome.wi.mit.edu/cgi-bin/primer/primer3_www.cgi) that would amplify ca. 500 base pair fragments in the reference strains (Table 1).

### **1.3. RD-PCR analysis**

5 Reactions were performed in 96 well plates and contained per reaction 1.25 µl of 10 x PCR buffer (600mM Tris HCl pH 8.8, 20 mM MgCl<sub>2</sub>, 170 mM (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>, 100 mM β-mercaptoethanol), 1.25 µl 20mM nucleotide mix, 50 nM of each primer, 1-10 ng of template DNA, 10% DMSO, 0.2 units *Taq* polymerase (Gibco-BRL) and sterile distilled water to 12.5 µl. Thermal cycling was performed on a PTC-100 amplifier (MJ Inc.) with an initial  
10 denaturation step of 90 seconds at 95°C, followed by 35 cycles of 30 seconds at 95°C, 1 min at 58°C, and 4 min at 72°C.

### **1.4. Sequencing of junction regions (RDs, TbD1,) *katG*, *gyrA*, *oxyR* and *pncA* genes**

PCR products were obtained as described above, using primers listed in Table 1.

15 For primer elimination, 6 µl PCR product was incubated with 1 unit of Shrimp Alkaline phosphatase (USB), 10 units of exonuclease I (USB), and 2 µl of 5 x buffer (200mM Tris HCl pH 8.8, 5mM MgCl<sub>2</sub>) for 15 min at 37°C and then for 15 min at 80°C. To this reaction mixture 2 µl of Big Dye sequencing mix (Applied Biosystems), 2 µl (2µM) of primer and 3 µl of 5 x buffer (5mM MgCl<sub>2</sub>, 200mM Tris HCl pH 8.8) were added and 35  
20 cycles (96°C for 30 sec; 56°C for 15 sec; 60°C for 4 min) performed in a thermocycler (MJ-research Inc., Watertown, MA). DNA was precipitated using 80 µl of 76% ethanol, centrifuged, rinsed with 70% ethanol, and dried. Reactions were dissolved in 2 µl of formamide/EDTA buffer, denatured and loaded onto 48 cm, 4 % polyacrylamide gels and electrophoresis performed on 377 automated DNA sequencers (Applied Biosystems) for 10  
25 to 12 h. Alternatively, reactions were dissolved in 0.3 mM EDTA buffer and subjected to automated sequencing on a 3700 DNA sequencer (Applied Biosystems). Reactions generally gave between 500-700 bp of unambiguous sequence.

### **1.5. Accession Numbers**

30 The sequence of the TbD1 region from the ancestral *M. tuberculosis* strain No. 74 (Ref. 8) containing genes *mmpS6* and *mmpL6* was deposited in the EMBL database under accession No. AJ426486. Sequences bordering RD4, RD7, RD8, RD9 and RD10 in BCG are available under accession numbers AJ003103, AJ007301, AJ131210, Y18604, and AJ132559, respectively.

## 2. EXPERIMENTAL DATA:

The distribution of 20 variable regions resulting from insertion-deletion events in the genomes of the tubercle bacilli has been evaluated in a total of 100 strains of *Mycobacterium tuberculosis*, *M. africanum*, *M. canettii*, *M. microti* and *M. bovis*. This approach showed that the majority of these polymorphisms did not occur independently in the different strains of the *M. tuberculosis* complex but, rather, result from ancient, irreversible genetic events in common progenitor strains. Based on the presence or absence of an *M. tuberculosis* specific deletion (TbD1), *M. tuberculosis* strains can be divided into ancestral and "modern" strains, the latter comprising representatives of major epidemics like the Beijing, Haarlem and African *M. tuberculosis* clusters. Furthermore, successive loss of DNA, reflected by RD9 and other subsequent deletions, was identified for an evolutionary lineage represented by *M. africanum*, *M. microti* and *M. bovis* that diverged from the progenitor of the present *M. tuberculosis* strains before TbD1 occurred. These findings contradict the often-presented hypothesis that *M. tuberculosis*, the etiological agent of human tuberculosis evolved from *M. bovis*, the agent of bovine disease. *M. canettii* and ancestral *M. tuberculosis* strains lack none of these deleted regions and therefore appear to be direct descendants of tubercle bacilli that existed before the *M. africanum*→*M. bovis* lineage separated from the *M. tuberculosis* lineage. This suggests that the common ancestor of the tubercle bacilli resembled *M. tuberculosis* or *M. canettii* and could well have been a human pathogen already.

The mycobacteria grouped in the *M. tuberculosis* complex are characterized by 99.9% similarity at the nucleotide level and identical 16S rRNA sequences (1, 2) but differ widely in terms of their host tropisms, phenotypes and pathogenicity. Assuming that they are all derived from a common ancestor, it is intriguing that some are exclusive human (*M. tuberculosis*, *M. africanum*, *M. canettii*) or rodent pathogens (*M. microti*) whereas others have a wide host spectrum (*M. bovis*). What was the genetic organization of the last common ancestor of the tubercle bacilli and in which host did it live? Which genetic events may have contributed to the fact that the host spectrum is so different and often specific? Where and when did *M. tuberculosis* evolve? Answers to these questions are important for a better understanding of the pathogenicity and the global epidemiology of tuberculosis and may help to anticipate future trends in the spread of the disease.

Because of the unusually high degree of conservation in their housekeeping genes it has been suggested that the members of the *M. tuberculosis* complex underwent an evolutionary bottleneck at the time of speciation, estimated to have occurred roughly 15,000



– 20,000 years ago (2). It also has been speculated that *M. tuberculosis*, the most widespread etiological agent of human tuberculosis has evolved from *M. bovis*, the agent of bovine tuberculosis, by specific adaptation of an animal pathogen to the human host (3). However, both hypotheses were proposed before the whole genome sequence of *M. tuberculosis* (4) was available and before comparative genomics uncovered several variable genomic regions in the members of the *M. tuberculosis* complex. Differential hybridization arrays identified 14 regions (RD1 –14) ranging in size from 2 to 12.7 kb that were absent from BCG Pasteur relative to *M. tuberculosis* H37Rv (5, 6). In parallel, six regions, RvD1–5, and TbD1, that were absent from the *M. tuberculosis* H37Rv genome relative to other members of the *M. tuberculosis* complex were revealed by comparative genomics approaches employing pulsed-field gel electrophoresis (PFGE) techniques (5, 7) and *in silico* comparisons of the near complete *M. bovis* AF2122/97 genome sequence and the *M. tuberculosis* H37Rv sequence.

In the present study the inventors have analyzed the distribution of these 20 variable regions situated around the genome (Table 1) in a representative and diverse set of 100 strains belonging to the *M. tuberculosis* complex. The strains were isolated from different hosts, from a broad range of geographic origins, and exhibit a wide spectrum of typing characteristics like IS6110 and spoligotype hybridization patterns or variable-number tandem repeats of mycobacterial interspersed repetitive units (MIRU-VNTR) (8, 9). The inventors have found striking evidence that deletion of certain variable genomic regions did not occur independently in the different strains of the *M. tuberculosis* complex and, assuming that there is little or no recombination of chromosomal segments between the various lineages of the complex, this allows the inventors to propose a completely new scenario for the evolution of the *M. tuberculosis* complex and the origin of human tuberculosis.

#### **Variable genomic regions and their occurrence in the members of the *M. tuberculosis* complex.**

The PCR screening assay for the 20 variable regions (Table 1) within 46 *M. tuberculosis*, 14 *M. africanum*, 5 *M. canettii*, 5 *M. microti*, 28 *M. bovis* and 2 BCG strains employed oligonucleotides internal to known RDs and RvDs, as well as oligonucleotides flanking these regions (Table 1). This approach generated a large data set that was robust, highly reliable, and internally controlled since PCR amplicons obtained with the internal

primer pair correlated with the absence of an appropriately sized amplicon with the flanking primer-pair, and *vice-versa*.

According to the conservation of junction sequences flanking the variable regions three types of regions were distinguished, each having different importance as an evolutionary marker. The first type included mobile genetic elements, like the prophages phiRv1 (RD3) and phiRv2 (RD11) and insertion sequences IS1532 (RD6) and IS6110 (RD5), whose distribution in the tubercle bacilli was highly divergent (Table 2). The second type of deletion is mediated by homologous recombination between adjacent IS6110 insertion elements resulting in the loss of the intervening DNA segment (RvD2, RvD3, RvD4, and RvD5 (7)) and is variable from strain to strain (Table 2).

The third type includes deletions whose bordering genomic regions typically do not contain repetitive sequences. Often this type of deletion occurred in coding regions resulting in the truncation of genes that are still intact in other strains of the *M. tuberculosis* complex. The exact mechanism leading to this type of deletion remains obscure, but possibly rare strand slippage errors of DNA polymerase may have contributed to this event. As shown in detail below, RD1, RD2, RD4, RD7, RD8, RD9, RD10, RD12, RD13, RD14, and TbD1 are representatives of this third group whose distribution among the 100 strains allows us to propose an evolutionary scenario for the members of the *M. tuberculosis* complex, that identified *M. tuberculosis* and/or *M. canettii* as most closely related to the common ancestor of the tubercle bacilli.

### 2.1. *M. tuberculosis* strains:

Investigation of the 46 *M. tuberculosis* strains by deletion analysis revealed that most RD regions were present in all *M. tuberculosis* strains tested (Table 2). Only regions RD3 and RD11, corresponding to the two prophages phiRv1 and phiRv2 of *M. tuberculosis* H37Rv (4), RD6 containing the insertion sequence IS1532, and RD5 that is flanked by a copy of IS6110 (5) were absent in some strains. This is an important observation as it implies that *M. tuberculosis* strains are highly conserved with respect to RD1, RD2, RD4, RD7, RD8, RD9, RD10, RD12, RD13, and RD14, and that these RDs represent regions that can differentiate *M. tuberculosis* strains independent of their geographical origin and their typing characteristics from certain other members of the *M. tuberculosis* complex. Furthermore, this suggests that these regions may be involved in the host specificity of *M. tuberculosis*.

In contrast, the presence or absence of RvD regions in *M. tuberculosis* strains was

variable. The region which showed the greatest variability was RvD2, since 18 from 46 tested *M. tuberculosis* strains did not carry the RvD2 region. Strains with a high copy number of IS6110 (>14) missed regions RvD2 to RvD5 more often than strains with only a few copies. As an example, all six tested strains belonging to the Beijing cluster (8) lacked regions RvD2 and RvD3. This is in agreement with the proposed involvement of recombination of two adjacent copies of IS6110 in this deletion event (7).

However, the most surprising finding concerning the RvD regions was that TbD1 was absent from 40 of the tested *M. tuberculosis* strains (87 %), including representative strains from major epidemics such as the Haarlem, Beijing and Africa clusters (8). To accentuate this result we named this region "*M. tuberculosis* specific deletion 1" (TbD1). *In silico* sequence comparison of *M. tuberculosis* H37Rv with the corresponding section in *M. bovis* AF2122/97 revealed that in *M. bovis* this locus comprises two genes encoding membrane proteins belonging to a large family, whereas in *M. tuberculosis* H37Rv one of these genes (*mmpS6*) was absent and the second was truncated (*mmpL6*). Unlike the RvD2-RvD5 deletions, the TbD1 region is not flanked by a copy of IS6110 in *M. tuberculosis* H37Rv, suggesting that insertion elements were not involved in the deletion of the 2153 bp fragment. To further investigate whether the 40 *M. tuberculosis* strains lacking the TbD1 region had the same genomic organization of this locus as *M. tuberculosis* H37Rv, we amplified the TbD1-junction regions of the various strains by PCR using primers flanking the deleted region (Table 1). This approach showed that the size of the amplicons obtained from multiple strains was uniform (Fig. 1) and subsequent sequence analysis of the PCR products revealed that in all tested TbD1-deleted strains the sequence of the junction regions was identical to that of *M. tuberculosis* H37Rv (Fig.2). The perfect conservation of the junction sequences in TbD1-deleted strains of wide geographical diversity suggests that the genetic event which resulted in the deletion occurred in a common progenitor. However, six *M. tuberculosis* strains, all characterized by very few or no copies of IS6110 and spoligotypes that resembled each other (Fig. 3) still had the TbD1 region present. Interestingly, these six strains were also clustered together by MIRU-VNTR analysis (9).

Analysis of partial gene sequences of *oxyR*, *pncA*, *katG*, and *gyrA* which have been described as variable between different tubercle bacilli (2, 11, 12, 13) revealed that all tested *M. tuberculosis* strains showed *oxyR* and *pncA* partial sequences typical for *M. tuberculosis* (*oxyR* - nucleotide 285 (*oxyR*<sup>285</sup>):G, *pncA* - codon 57 (*pncA*<sup>57</sup>: CAC ). Based on the *katG* codon 463 (*katG*<sup>463</sup>) and *gyrA* codon 95 (*gyrA*<sup>95</sup>) sequence polymorphism, Sreevatsan and colleagues (2) defined three groups among the tubercle bacilli, group 1 showing *katG*<sup>463</sup>

CTG (Leu), *gyrA*<sup>95</sup> ACC (Thr), group 2 exhibiting *katG*<sup>463</sup> CGG (Arg), *gyrA*<sup>95</sup> ACC (Thr), and group 3 showing *katG*<sup>463</sup> CGG (Arg), *gyrA*<sup>95</sup> AGC (Ser). According to this scheme, in our study 16 of the 46 tested *M. tuberculosis* strains belonged to group 1, whereas 27 strains belonged to group 2 and only 3 isolates to group 3. From the 40 strains that were deleted for region TbD1, 9 showed characteristics of group 1, including the strains belonging to the Beijing cluster, 28 of group 2, including the strains from the Haarlem and Africa clusters and 3 of group 3, including H37Rv and H37Ra. Most interestingly, all six *M. tuberculosis* strains where the TbD1 region was not deleted, contained a leucine (CTG) at *katG*<sup>463</sup>, which was described as characteristic for ancestral *M. tuberculosis* strains (group 1) (2). As shown in Figure 4, this suggests that during the evolution of *M. tuberculosis* the *katG* mutation at codon 463 CTG (Leu) → CGG (Arg) occurred in a progenitor strain that had region TbD1 deleted. This proposal is supported by the finding that strains belonging to group 1 may or may not have deleted region TbD1, whereas all 30 strains belonging to groups 2 and 3 lacked TbD1 (Fig. 4). Furthermore, all strains of groups 2 and 3 characteristically lacked spacer sequences 33-36 in the direct repeat (DR) region (Fig. 3). It appears that such spacers may be lost but not gained (14). Therefore, TbD1 deleted strains will be referred to hereafter as “modern” *M. tuberculosis* strains.

## 2.2. *M. canettii*:

*M. canettii* is a very rare smooth variant of *M. tuberculosis*, isolated usually from patients from, or with connection to, Africa. Although it shares identical 16S rRNA sequences with the other members of the *M. tuberculosis* complex, *M. canettii* strains differ in many respects including polymorphisms in certain house-keeping genes, IS1081 copy number, colony morphology, and the lipid content of the cell wall (15, 16). Therefore, we were surprised to find that in *M. canettii* all the RD, RvD, and TbD1 regions except the prophages (phiRv1, phiRv2) were present. In contrast, we identified a region (RD<sup>can</sup>) being specifically absent from all five *M. canettii* strains that partially overlapped RD12 (Fig. 4).

The conservation of the RD, RvD, and TbD1 regions in the genome of *M. canettii* in conjunction with the many described and observed differences suggest that *M. canettii* diverged from the common ancestor of the *M. tuberculosis* complex before RD, RvD and TbD1 occurred in the lineages of tubercle bacilli (Fig. 4). This hypothesis is supported by the finding that *M. canettii* was shown to carry 26 unique spacer sequences in the direct repeat region (14), that are no longer present in any other member of the *M. tuberculosis* complex. Therefore, *M. canettii* represents a fascinating tubercle bacillus, whose detailed genomic

analysis may reveal further insights into the evolution of the *M. tuberculosis* complex.

### 2.3. *M. africanum*:

The isolates designated as *M. africanum* studied here originate from West and East-African sources. 11 strains were isolated in Sierra Leone, Nigeria and Guinea and 2 strains in Uganda. One strain comes from the Netherlands.

For the 11 West African isolates, RD analysis indicated that these strains all lack the RD9 region containing *cobL*. Sequence analysis of the RD9 junction region showed that the genetic organization of this locus in West African strains was identical to that of *M. bovis* and *M. microti* in that the 5' part of *cobL* as well as the genes Rv2073c and Rv2074c were absent. In addition, six strains (2 from Sierra Leone, 4 from Guinea) also lacked RD7, RD8 and RD10 (Table 2). The junction sequences bordering RD7, RD8 and RD10, like those for RD9, were identical to those of *M. bovis* and *M. microti* strains. As regards the two prophages phiRv1 and phiRv2, the West African strains all contained phiRv2, whereas phiRv1 was absent. No variability was seen for the RvD regions. RvD1-RvD5 and TbD1 were present in all tested West African strains. This shows that *M. africanum* prevalent in West Africa can be differentiated from "modern" *M. tuberculosis* by at least two variable genetic markers, namely the absence of region RD9 and the presence of region TbD1.

In contrast, for East African *M. africanum* and for the isolate from the Netherlands, no genetic marker was found which could differentiate them from *M. tuberculosis* strains. With the exception of prophage phiRv1 (RD3) the 3 strains from Uganda and the Netherlands did not exhibit any of the RD deletions, but lacked the TbD1 region, as do "modern" *M. tuberculosis* strains. The absence of the TbD1 region was also confirmed by sequence analysis of the TbD1 junction region, which was found to be identical to that of TbD1 deleted *M. tuberculosis* strains. These results indicate a very close genetic relationship of these strains to *M. tuberculosis* and suggest that they should be regarded as *M. tuberculosis* rather than *M. africanum* strains.

### 2.4. *M. microti*:

*M. microti* strains were isolated in the 1930's from voles (17) and more recently from immuno-suppressed patients (18). These strains are characterized by an identical, characteristic spoligotype, but differ in their IS6110 profiles. Both, the vole and the human isolates, lacked regions RD7, RD8, RD9, and RD10 as well as a region that is specifically deleted from *M. microti* (RD<sup>mic</sup>). RD<sup>mic</sup> was revealed by a detailed comparative genomics

study of *M. microti* isolates (19) using clones from a *M. microti* Bacterial Artificial Chromosome (BAC) library. RD<sup>mic</sup> partially overlaps RD1 from BCG (data not shown). Furthermore, vole isolates missed part of the RD5 region, whereas this region was present in the human isolate. As the junction region of RD5 in *M. microti* was different to that in BCG (data not shown), RD5 was not used as an evolutionary marker.

### 2.5. *M. bovis* and *M. bovis* BCG:

*M. bovis* has a very large host spectrum infecting many mammalian species, including man. The collection of *M. bovis* strains that was screened for the RD and RvD regions consisted of 2 BCG strains and 18 "classical" *M. bovis* strains generally characterized by only one or two copies of IS6110 from bovine, llama and human sources in addition to three goat isolates, three seal isolates, two oryx isolates, and two *M. bovis* strains from humans that presented a higher number of IS6110 copies.

Excluding prophages, the distribution of RDs allowed us to differentiate five main groups among the tested *M. bovis* strains. The first group was formed by strains that lack RD7, RD8, RD9, and RD10. Representatives of this group are three seal isolates and two human isolates containing between three and five copies of IS6110 (data not shown). Two oryx isolates harboring between 17 and 20 copies of IS6110 formed the second group that lacked parts of RD5 in addition to RD7-RD10, and very closely resembled the *M. microti* isolates. However, they did not show RD<sup>mic</sup>, the deletion characteristic of *M. microti* strains (data not shown). Analysis of partial *oxyR* and *pncA* sequences from strains belonging to groups one and two, showed sequence polymorphisms characteristic of *M. tuberculosis* strains (*oxyR*<sup>285</sup>: G, *pncA*<sup>57</sup>: CAC, Ref. 12, 13).

Group three consists of goat isolates that lack regions RD5, RD7, RD8, RD9, RD10, RD12, and RD13. As previously described by Aranaz and colleagues, these strains exhibited an adenosine at position 285 of the *oxyR* pseudogene that is specific for "classical" *M. bovis* strains whereas the sequence of the *pncA*<sup>57</sup> polymorphism was identical to that in *M. tuberculosis* (20). This is in good agreement with our results from sequence analysis (Table 2) and the finding that except for RD4, the goat isolates displayed the same deletions as "classical" *M. bovis* strains. Taken together, this suggests that the *oxyR*<sup>285</sup> mutation (G → A) occurred in *M. bovis* strains before RD4 was lost. Interestingly, the most common *M. bovis* strains ("classical" *M. bovis* (21)), isolated from cattle from Argentina, the Netherlands, the UK and Spain, as well as from humans (e. g. multi-drug resistant *M. bovis*

from Spain (22)) showed the greatest number of RD deletions and appear to have undergone the greatest loss of DNA relative to other members of the *M. tuberculosis* complex. These lacked regions RD4, RD5, RD6, RD7, RD8, RD9, RD10, RD12 and RD13, confirming results obtained with reference strains (5, 6). These strains together with the two BCG strains  
 5 were the only ones that showed the *pncA*<sup>57</sup> polymorphism GAC (Asp) in addition to the *oxyR*<sup>285</sup> mutation (G → A) characteristic of *M. bovis*. Analysis of BCG strains indicate that BCG lacked the same RD regions as “classical” *M. bovis* strains in addition to RD1, RD2 and RD14 which apparently occurred during and after the attenuation process (Fig. 4) (6, 23).

10 In contrast to RDs, the RvD regions were highly conserved in the *M. bovis* strains. With the exception of the two IS6110-rich oryx isolates, that lacked RvD2, RvD3 and RvD4, all other strains had the five RvD regions present. It is particularly noteworthy that TbD1 was present in all *M. bovis* strains.

However, except for the two human isolates, containing between three and five copies  
 15 of IS6110 from group 1, strains designated as *M. bovis* showed a single nucleotide polymorphism in the TbD1 region at codon 551 (AAG) of the *mmpL6* gene, relative to *M. canettii*, *M. africanum* and ancestral *M. tuberculosis* strains, which are characterized by codon AAC. Even the strains isolated from seals and from oryx with *oxyR* or *pncA* loci like those of *M. tuberculosis* and with fewer deleted regions than the classical *M. bovis* strains,  
 20 showed the *mmpL6*<sup>551</sup>AAG polymorphism typical for *M. bovis* and *M. microti* (Table 2, Fig. 4). As such, this polymorphism could serve as a very useful genetic marker for the differentiation of strains that lack RD7, RD8, RD9, and RD10 and have been classified as *M. bovis* or *M. africanum*, but may differ from other strains of the same taxon.

## 25 3. DISCUSSION

### 3.1. Origin of human tuberculosis

For many years, it was thought that human tuberculosis evolved from the bovine disease by adaptation of an animal pathogen to the human host (3). This hypothesis is based  
 30 on the property of *M. tuberculosis* to be almost exclusively a human pathogen, whereas *M. bovis* has a much broader host range. However, the results from this study unambiguously show that *M. bovis* has undergone numerous deletions relative to *M. tuberculosis*. This is confirmed by the preliminary analysis of the near complete genome sequence of *M. bovis* AF2122/97, a “classical” *M. bovis* strain isolated from cattle, which revealed no new gene

clusters that were confined specifically to *M. bovis*. This indicates that the genome of *M. bovis* is smaller than that of *M. tuberculosis* (24). It seems plausible that *M. bovis* is the final member of a separate lineage represented by *M. africanum* (RD9), *M. microti* (RD7, RD8, RD9, RD10) and *M. bovis* (RD4, RD5, RD7, RD8, RD9, RD10, RD12, RD13) (25) that  
 5 branched from the progenitor of *M. tuberculosis* isolates. Successive loss of DNA may have contributed to clonal expansion and the appearance of more successful pathogens in certain new hosts.

Whether the progenitor of extant *M. tuberculosis* strains was already a human pathogen when the *M. africanum* → *M. bovis* lineage separated from the *M. tuberculosis*  
 10 lineage is a subject for speculation. However, we have two reasons to believe that this was the case. Firstly, the six ancestral *M. tuberculosis* strains (TbD1<sup>+</sup>, RD9<sup>+</sup>) (Fig.3) that resemble the last common ancestor before the separation of *M. tuberculosis* and *M. africanum* are all human pathogens. Secondly, *M. canettii*, which probably diverged from the common ancestor of today's *M. tuberculosis* strains prior to any other known member of the  
 15 *M. tuberculosis* complex is also a human pathogen. Taken together, this means that those tubercle bacilli, which are thought to most closely resemble the progenitor of *M. tuberculosis* are human and not animal pathogens. It is also intriguing that most of these strains were of African or Indian origin (Fig. 3). It is likely that these ancestral strains predominantly originated from endemic foci (15, 26), whereas "modern" *M. tuberculosis* strains that have  
 20 lost TbD1 may represent epidemic *M. tuberculosis* strains that were introduced into the same geographical regions more recently as a consequence of the worldwide spread of the tuberculosis epidemic.

### 3.2. The evolutionary timescale of the *M. tuberculosis* complex

25 Because of the high sequence conservation in housekeeping genes, Sreevatsan *et al.* previously hypothesized that the tubercle bacilli encountered a major bottleneck 15,000 – 20,000 years ago (2). As the conservation of the TbD1 junction sequence in all tested TbD1 deleted strains suggests descentance from a single clone, the TbD1 deletion is a perfect indicator that "modern" *M. tuberculosis* strains that account for the vast majority of today's  
 30 tuberculosis cases definitely underwent such a bottleneck and then spread around the world.

As described in detail in the results section, our analysis showed that the *katG*<sup>463</sup> CTG→CGG and the subsequent *gyrA*<sup>95</sup> ACC →AGC mutations, that were used by Sreevatsan and colleagues to designate groups 2 and 3 of their proposed evolutionary pathway of the tubercle bacilli (2), occurred in a lineage of *M. tuberculosis* strains that had



already lost TbD1 (Fig.4). Although deletions are more stable markers than point mutations, which may be subject to reversion, a perfect correlation of deletion and point mutation data was found for the tested strains.

This information, together with results from a recent study by Fletcher and colleagues (27), who have shown that *M. tuberculosis* DNAs amplified from naturally mummified Hungarian villagers from the 18<sup>th</sup> and 19<sup>th</sup> century belonged to *katG*<sup>463</sup>/*gyrA*<sup>95</sup> groups 2 and 3, suggests that the TbD1 deletion occurred in the lineage of *M. tuberculosis* before the 18<sup>th</sup> century. This could mean that the dramatic increase of tuberculosis cases later in the 18<sup>th</sup> century in Europe mainly involved “modern” *M. tuberculosis* strains. In addition, it shows that tuberculosis was caused by *M. tuberculosis* and not by *M. bovis*, a fact which is also described for cases in rural medieval England (28).

There is good evidence that mycobacterial infections occurred in man several thousand years ago. We know that tuberculosis occurred in Egypt during the reign of the pharaohs because spinal and rib lesions pathognomonic of tuberculosis have been identified in mummies from that period (29). Identification of acid fast bacilli as well as PCR amplification of IS6110 from Peruvian mummies (30) also suggest that tuberculosis existed in pre-Columbian societies of Central and South America. To estimate when the TbD1 bottleneck occurred, it would now be very interesting to know whether the Egyptian and South American mummies carried *M. tuberculosis* DNA that had TbD1 deleted or not.

The other major bottleneck, which seems to have occurred for members of the *M. africanum* → *M. microti* → *M. bovis* lineage is reflected by RD9 and the subsequent RD7, RD8 and RD10 deletions (Fig. 4). These deletions seem to have occurred in the progenitor of tubercle bacilli that - today - show natural host spectra as diverse as humans in Africa, voles on the Orkney Isles (UK), seals in Argentina, goats in Spain, and badgers in the UK. For this reason it is difficult to imagine that spread and adaptation of RD9-deleted bacteria to their specific hosts could have appeared within the postulated 15,000 – 20,000 years of speciation of the *M. tuberculosis* complex.

However, more insight into this matter could be gained by RD analysis of ancient DNA samples, e. g. mycobacterial DNA isolated from a 17,000 year old bison skeleton (31). The mycobacterium whose DNA was amplified showed a spoligotype that was most closely related to patterns of *M. africanum* and could have been an early representative of the lineage *M. africanum* → *M. bovis*. With the TbD1 and RD9 junction sequences that we supply here, PCR analyses of ancient DNAs should enable very focused studies to be undertaken to learn more about the timescale within which the members of the *M.*

*tuberculosis* complex have evolved.

### 3.3. Concluding comments

Our study provides an overview of the diversity and conservation of variable regions in a broad range of tubercle bacilli. Deletion analysis of 100 strains from various hosts and countries has identified some evolutionarily "old" *M. canettii*, *M. tuberculosis* and *M. africanum* strains, most of them of African origin, as well as "modern" *M. tuberculosis* strains, the latter including representatives from major epidemic clusters like Beijing, Haarlem and Africa. The use of deletion analysis in conjunction with molecular typing and analysis of specific mutations was shown to represent a very powerful approach for the study of the evolution of the tubercle bacilli and for the identification of evolutionary markers. In a more practical perspective, these regions, primarily RD9 and TbD1 but also RD1, RD2, RD4, RD7, RD8, RD10, RD12 and RD13 represent very interesting candidates for the development of powerful diagnostic tools for the rapid and unambiguous identification of members of the *M. tuberculosis* complex (32). This genetic approach for differentiation can now be used to replace the often confusing traditional division of the *M. tuberculosis* complex into rigidly defined subspecies.

Moreover, functional analyses will show whether the TbD1 deletion confers some selective advantage to "modern" *M. tuberculosis*, or whether other circumstances contributed to the pandemic of the TbD1 deleted *M. tuberculosis* strains.

#### EXAMPLE 4

The members of the *M. tuberculosis* complex share an unusually high degree of conservation such that the commercially-available nucleic acid probes and amplification assays cannot differentiate these organisms. In addition conventional identification methods are often ambiguous, cumbersome and time consuming because of the slow growth of the organisms.

In the present invention the inventors, by a deletion analysis, solve the problem faced by clinical mycobacteriology laboratories for differentiation within the *M. tuberculosis* complex.

This approach allows to perform a diagnostic on a biological fluid by using at least three markers including TBD1. The following table 3 illustrates such a combination sufficient to realize the distinction between the members of the *M. tuberculosis* complex.

MYCOBACTERIUM STRAIN	MARKERS		
	RD4	RD9	TbD1
M. bovis BCG	-	-	+
M. bovis	-	-	+
M. africanum	+	-	+
M. tuberculosis	+	+	-
M. tuberculosis ancêtre	+	+	+
M. canettii	+	+	+

Table 3

Beside TbD1 marker, preferably at least 2 other markers should be used. Examples of such  
5 additional markers available in the literature are listed in the following table 1.

Supplemental data:

**Table 1: RD, RvD and TbD1 regions and selected primers**

Region absent from BCG	Gene	Size (kb)	Internal Primerpair	Flanking primers or 2 <sup>nd</sup> internal * primerpair
RD1	Rv3871-Rv3879c	9.5	RD1in-Rv3878F GTC AGC CAA GTC AGG CTA CC RD1in-Rv3878R CAA CGT TGT GGT TGT TGA GG	RD1-flank.left GAA ACA GTC CCC AGC AGG T RD1-flank.right TTC AAC GGG TTA CTG CGA AT
RD2	Rv1978-Rv1988	10.8	RD2-Rv1979.int.F TAT AGC TCT CGG CAG GTT CC RD2-Rv1979-int.R ATC GGC ATC TAT GTC GGT GT	RD2-flank.F CTC GAC CGC GAC GAT GTG C RD2-flank.R CCT CGT TGT CAC CGC GTA TG
RD3*	Rv1573-Rv1586c	9.2	RD3-Rv1586.int.F TTA TCT TGG CGT TGA CGA TG RD3-Rv1586.int.R CAT ATA AGG GTG CCC GCT AC	RD3-int-REP.F CTG ACG TCG TTG TCG AGG TA* RD3-int-REP.R GTA CCC CCA GGC GAT CTT*
RD4	Rv1505c-Rv1516c	12.7	RD4-Rv1516.int.F CAA GGG GTA TGA GGT TCA CG RD4-Rv1516.int.R CGG TGA TTC GTG ATT GAA CA	RD4-flank.F CTC GTC GAA GGC CAC TAA AG RD4-flank.R AAG GCG AAC AGA TTC AGC AT
RD5*	Rv2346c-Rv2353c	9.0	RD5A-Rv2348.int.F AAT CAC GCT GCT GCT ACT CC RD5A-Rv2348.int.R GTG CTT TTG CCT CTT GGT C	RD5B-plcA.int.F CAA GTT GGG TCT GGT CGA AT RD5B-plcA.int.R GCT ACC CAA GGT CTC CTG GT
RD6*	Rv3425-Rv3428c	4.9	RD6-IS1532F CAG CTG GTG AGT TCA AAT GC RD6-IS1532R CTC CCG ACA CCT GTT CGT	ND ND
RD7	Rv1964-Rv1977	12.7	RD7-Rv1976.int.F TGG ATT GTC GAC GGT ATG AA RD7-Rv1976.int.R GGT CGA TAA GGT CAC GGA AC	RD7-flank.F GGT AAT CGT GGC CGA CAA G RD7-flank.R CAG CTC TTC CCC TCT CGA C
RD8	<i>ephA-lpqG</i>	5.9	RD8-ephA.F GGT GTG ATT TGG TGA GAC GAT G	RD8-flank.F CAA TCA GGG CTG TGC TAA CC

RD9	cobL-Rv2075	2.0	RD8-ephA.R	RD8-flank.R
			AGT TCC TCC TGA CTA ATC CAG GC	CGA CAG TTG TGC GTA CTG GT
			RD9-intF	RD9-flankF
			CGA TGG TCA ACA CCA CTA CG	GTG TAG GTC AGC CCC ATC C
RD10	Rv0221-Rv0223	1.9	RD9-intR	RD9-flankR
			CTG GAC CTC GAT GAC CAC TC	GCC CAA CAG CTC GAC ATC
			RD10-intF	RD10-flankF
			GTA ACC GCT TCA CCG GAA T	CTG CAA CCA TCC GGT ACA C
RD11	Rv2645-Rv2695c	11.0	RD10-intR	RD10-flankR
			GTC AAC TCC ACG GAA AGA CC	GTC ATG AAC GCC GGA CAG
			RD11-Rv2646F	RD11-fla-F
			CGG CAG CTA GAC GAC CTC	TCA CAT AGG GGC TGC GAT AG
RD12	sseC-Rv3121	2.8	RD11-Rv2646R	RD11-fla-R
			AAC GTG CTG CGA TAG GTT TT	AGA GGA ACC TTT CGG TGG TT
			RD12-Rv3120.int.F	RD12-flank.F
			GAA ATA CGA GTG CGC TGA CC	GCC ATC AAC GTC AAG AAC CT
RD13	Rv1255c-Rv1257c	3.0	RD12-Rv3120.int.R	RD12-flank.R
			CTC TGA ACC ATC GGT GTC G	CGG CCA GGT AAC AAG GAG T
			RD13intF	RD13-flank.F
			GGA TGT CAC TCG GAA CGG CA	CGA TGG TGT TTC TTG GTG AG
RD14	Rv1765c-Rv1773c	9.0	RD13intR	RD13-flank.R
			CAC CGG GCT GAT CGA GCG A	GGA TCG GCT CAG TGA ATA CC
			RD14-Rv1769.int.F	RD14-flankF
			GTG GAG CAC CTT GAC CTG AT	TTG ATT CGC CAA CAA CTG AA
			RD14-Rv1769.int.R	RD14-flankR
			CGT CGA ATA CGA GTC GAA CA	GGG CTG GTT AGT GTC GAT TC
			Region missing from M. tuberculosis H37Rv	
			RvD1*	5.0
AGC GCG TCG AAC ACC GGC	GAG CCA CTC CGA TGT TGA CT			
RvD1-int1R	RvD1-int2.R			
CCT GAA TCC GCG CAA TTC CAT	CAC GCG AAC CCT ACC TAC AT			
RvD2*	plcD	5.1	RvD2-int1F	RvD2-int2F
			GTT CTC CTG TCG AAC CTC CA	GGA CGG TGA CGG TAT TTG TC
			RvD2-int1R	RvD2-int2R
			ACT TCA CCG GTT TCA TCT CG	TCG CCA ACT TCT ATG GAC CT
RvD3		1.0	RvD3-intF	RvD3-flank.F
			ATC GAT CAG GTC GTC AAT GC	AAA CCA TGC AGC GTC TGC CA
			RvD3-intR	RvD3-flankR
			ACG CCA CCA TCA AGA TCC	GCG TTT CTG CGT CTG GTT GA

RvD4*	PPE gene	0.8	RvD4-intF-PPE GGT TGC CAA CGT TAC CGA TGC	ND
			RvD4-intR-PPE CCG GTG GTG GTG GCG GCT	ND
RvD5	<i>moa</i>	4.0	RvD5intF GGG TTC ACG TTC ATT ACT GTT C	RvD5-flankF CCC ATC GTG GTC GTT CAC C
			RvD5intR CCT GCG CTT ATC TCT AGC GG	RvD5-flankR GTA CCC GCA CCA CCT GCT G
TbD1	<i>mmpL6</i>	2.1	TbD1intS.F CGT TCA ACC CCA AAC AGG TA	TbD1fla1-F CTA CCT CAT CTT CCG GTC CA
			TbD1intS.R AAT CGA ACT CGT GGA ACA CC	TbD1fla1-R CAT AGA TCC CGG ACA TGG TG
<b><i>katG</i>, <i>gyrA</i>, <i>oxyR</i>', <i>pncA</i> and <i>mmpL6</i> PCR and sequencing primers</b>				
<i>katG</i> <sup>463</sup>			<i>katG</i> -2154,225-PCR-F CTA CCA GCA CCG TCA TCT CA	<i>katG</i> -2154,872-SEQ-R ACA AGC TGA TCC ACC GAG AC
			<i>katG</i> -2155,157-PCR-R AGG TCG TAT GGA CGAACA CC	
<i>gyrA</i> <sup>95</sup>			<i>gyrA</i> -7,127-PCR-F GTT CGT GTG TTG CGT CAA GT	<i>gyrA</i> -7,461F CGG GTG CTC TAT GCA ATG TT
			<i>gyrA</i> - 8,312-PCR-R CAG CTG GGT GTG CTT GTA AA	
<i>oxyR</i> <sup>285</sup>			<i>oxyR</i> 2725,559F TAT GCG ATC AGG CGT ACT TG	<i>oxyR</i> -2726,024-SEQ-R CAA AGC AGT GGT TCA GCA GT
			<i>oxyR</i> -2726,024-PCR-R CAA AGC AGT GGT TCA GCA GT	
<i>pncA</i> <sup>57</sup>			<i>pncA</i> -2288,678-PCR-F ATC AGG AGC TGC AAA CCA AC	<i>pncA</i> - 2289,319-SEQ-R GGC GTC ATG GAC CCT ATA TC
			<i>pncA</i> - 2289,319-PCR-R GGC GTC ATG GAC CCT ATA TC	
<i>mmpL6</i> <sup>551</sup>			<i>mmpL</i> -seq5F GTA TCA GAG GGA CCG AGC AG	<i>mmpL</i> -seq5F GTA TCA GAG GGA CCG AGC AG
			TbD1fla1-R CAT AGA TCC CGG ACA TGG TG	

The RD nomenclature used in this table is based on that used by Brosch *et al.* (2000), (Ref. 25) and differs from that proposed by Behr and coworkers (1999), (Ref. 6). Primer sequences are shown in 5' →3' direction.

\* Regions where a second pair of internal primers was used rather than flanking primers, due to

5 flanking repetitive regions, and/or mobile genetic elements.







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## CLAIMS

1. An isolated or purified nucleic acid wherein said nucleic acid is selected from the group consisting of:
  - 5           a.       SEQ ID N°1;
  - b.       Nucleic acid having a sequence fully complementary to SEQ ID N°1.
  - c.       Nucleic acid fragment comprising at least 15 consecutive nucleotides of  
                    SEQ ID N°1;
  - d.       Nucleic acid having at least 90% sequence identity after optimal  
10           alignment with a sequence defined in a) or b);
  - e.       Nucleic acid that hybridizes under stringent conditions with the nucleic  
                    acid defined in a) or b);
  
2. A nucleic acid fragment according to claim 1 wherein said nucleic acid fragment is:
  - 15           - specifically deleted from the genome of *Mycobacterium tuberculosis*, excepted in  
            *Mycobacterium tuberculosis* strain having the mutation CTG -> CGG at codon 463 of gene  
            katG and/or *Mycobacterium tuberculosis* strain having no or very few IS6110 sequences  
            inserted in their genome; and,
  - present in the genome of *Mycobacterium africanum*, *Mycobacterium canetti*,  
20       *Mycobacterium microti*, *Mycobacterium bovis*, *Mycobacterium bovis BCG*.
  
3. A nucleic acid fragment according to claim 1 or 2 selected from the group consisting  
of:
  - a) SEQ ID N°4;
  - 25       b) Nucleic acid having a sequence fully complementary to SEQ ID N°4.
  - c) Nucleic acid fragment comprising at least 20 consecutive nucleotides of SEQ ID  
            N°4;
  - d) Nucleic acid having at least 90% sequence identity after optimal alignment with a  
            sequence defined in a) or b);
  - 30       e) Nucleic acid that hybridizes under stringent conditions with the nucleic acid defined  
            in a) or b);
  
4. The isolated or purified nucleic acid of claim 1 wherein said nucleic acid comprises at  
least a deletion of a nucleic acid fragment according to claim 2 or 3.

5. Isolated or purified polypeptides encoded by the nucleic acid of claims 1 to 4.

5 6. The polypeptide of claim 5 selected among polypeptide with sequence SEQ ID N° 5, N° 7, N° 9, N°11, and their fragments thereof.

7. Isolated or purified nucleic acid encoding the polypeptide according to claim 6.

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10 8. Isolated or purified nucleic acid according to claim 7 selected among:

- SEQ ID N° 4 encoding the polypeptide of SEQ ID N°5;
- SEQ ID N° 6 encoding the polypeptide of SEQ ID N°7;
- SEQ ID N° 8 encoding the polypeptide of SEQ ID N°9;
- SEQ ID N° 10 encoding the polypeptide of SEQ ID N°11;

15 and their fragments thereof.

9. A recombinant vector comprising a nucleic acids sequence selected among nucleic acids according to claims 1 to 4 , 7 and 8.

20 ... 10. The recombinant vector of claim 9 consisting of vector named X229 introduced into the recombinant *Escherichia coli* deposited with the CNCM on February 18<sup>th</sup>, 2002 under N° I-2799.

25 11. A recombinant cell comprising a nucleic acids sequence selected among nucleic acids according to claims 1 to 4 , 7 and 8, or a vector according to claims 9 and 10.

12. The recombinant cell according to claim 11 consisting of the *Escherichia coli* deposited with the CNCM on February 18<sup>th</sup>, 2002 under N° I-2799. --

30 13. A method for the discriminatory detection and identification of:

- *Mycobacterium tuberculosis* excepted *Mycobacterium tuberculosis* strain having the mutation CTG -> CGG at codon 463 of gene katG and/or excepted *Mycobacterium tuberculosis* strain having no or very few IS6110 sequences inserted in their genome; versus,

- *Mycobacterium africanum*, *Mycobacterium canetti*, *Mycobacterium microti*,  
*Mycobacterium bovis*, *Mycobacterium bovis BCG* in a biological sample,  
 comprising the following steps:

- a) isolation of the DNA from the biological sample to be analyzed or  
 5 production of a cDNA from the RNA of the biological sample,
- b) detection of the nucleic acid sequences of the mycobacterium present in said  
 biological sample,
- c) analysis for the presence or the absence of a nucleic acid fragment according  
 to claims 2 or 3.

10

14. The method as claimed in claim 13, in which the detection of the mycobacterial DNA  
 sequences is carried out using nucleotide sequences complementary to said DNA sequences.

15. The method as claimed in claim 13 or 14, in which the detection of the mycobacterial  
 15 DNA sequences is carried out by amplification of these sequences using primers.

16. The method as claimed in claim 15, in which the primers have a nucleotide sequence  
 chosen from the group comprising SEQ ID N°13, SEQ ID N°14, SEQ ID N°15, SEQ ID  
 N°16, SEQ ID N°17, SEQ ID N°18.

20

17. A method for the discriminatory detection and identification of:

- *Mycobacterium tuberculosis* excepted *Mycobacterium tuberculosis* strain having the  
 mutation CTG → CGG at codon 463 of gene *katG* and/or excepted *Mycobacterium*  
*tuberculosis* strain having no or very few IS6110 sequences inserted in their genome; versus,
- 25 - *Mycobacterium africanum*, *Mycobacterium canetti*, *Mycobacterium microti*,  
*Mycobacterium bovis*, *Mycobacterium bovis BCG* in a biological sample,  
 comprising the following steps:

- a) bringing the biological sample to be analyzed into contact with at least one  
 pair of primers as defined in claim 15 or 16, the DNA contained in the sample having been,  
 30 where appropriate, made accessible to the hybridization beforehand,
- b) amplification of the DNA of the mycobacterium,
- c) visualization of the amplification of the DNA fragments.

18. A kit for the discriminatory detection and identification of:

- *Mycobacterium tuberculosis* excepted *Mycobacterium tuberculosis* strain having the mutation CTG → CGG at codon 463 of gene katG and/or excepted *Mycobacterium tuberculosis* strain having no or very few IS6110 sequences inserted in their genome; versus,
- *Mycobacterium africanum*, *Mycobacterium canetti*, *Mycobacterium microti*,  
5 *Mycobacterium bovis*, *Mycobacterium bovis BCG* in a biological sample,  
in a biological sample comprising the following elements:

- a) at least one pair of primers as defined in claim 15 or 16,
- b) the reagents necessary to carry out a DNA amplification reaction,
- c) optionally, the necessary components which make it possible to verify or  
10 compare the sequence and/or the size of the amplified fragment.

19. The use of at least one pair of primers as defined in claim 15 or 16 for the amplification of a DNA sequence from *Mycobacterium tuberculosis* or *Mycobacterium africanum*, *Mycobacterium canetti*, *Mycobacterium microti*, *Mycobacterium bovis*,  
15 *Mycobacterium bovis BCG*, *Mycobacterium tuberculosis* having the mutation CTG → CGG at codon 463 of gene katG and/or having no or very few IS6110 sequences inserted in their genome.

20. A product of expression of all or part of the nucleic acid fragment as defined in claim 2 or 3 deleted from the genome of *Mycobacterium tuberculosis* and present in *Mycobacterium africanum*, *Mycobacterium canetti*, *Mycobacterium microti*, *Mycobacterium bovis*, *Mycobacterium bovis BCG*, *Mycobacterium tuberculosis* having the mutation CTG → CGG at codon 463 of gene katG and/or having no or very few IS6110 sequences inserted in their genome; or conversely.

21. A method for the discriminatory detection *in vitro* of antibodies directed against *Mycobacterium tuberculosis* or *Mycobacterium africanum*, *Mycobacterium canetti*, *Mycobacterium microti*, *Mycobacterium bovis*, *Mycobacterium bovis BCG*, *Mycobacterium Tuberculosis* having the mutation CTG → CGG at codon 463 of gene KatG and/or having no  
30 or very few IS6110 sequences inserted in their genome, in a biological sample, comprising the following steps:

- a) bringing the biological sample into contact with at least one product as defined in claim 20,
- b) detecting the antigen-antibody complex formed.

22. A method for the discriminatory detection of a vaccination with *Mycobacterium bovis* BCG or an infection by *Mycobacterium tuberculosis*, excepted *Mycobacterium Tuberculosis* strain having the mutation CTG → CGG at codon 463 of gene KatG and/or  
5 excepted *Mycobacterium Tuberculosis* strain having no or very few IS6110 sequences inserted in their genome in a mammal, comprising the following steps:

- a) preparation of a biological sample containing cells, more particularly cells of the immune system of said mammal and more particularly T cells,
- b) incubation of the biological sample of step a) with at least one product as  
10 defined in claim 20,
- c) detection of a cellular reaction indicating prior sensitization of the mammal to said product, in particular cell proliferation and/or synthesis of proteins such as gamma-interferon.

15 23. A kit for the *in vitro* diagnosis of *Mycobacterium tuberculosis* infection, excepted infection with *Mycobacterium tuberculosis* strain having the mutation CTG → CGG at codon 463 of gene katG and/or having no or very few IS6110 sequences inserted in their genome, in a mammal optionally vaccinated beforehand with *M. bovis* BCG comprising:

- a) a product as defined in claim 20,
- 20 b) where appropriate, the reagents for the constitution of the medium suitable for the immunological reaction,
- c) the reagents allowing the detection of the antigen-antibody complexes produced by the immunological reaction,
- d) where appropriate, a reference biological sample (negative control) free of  
25 antibodies recognized by said product,
- e) where appropriate, a reference biological sample (positive control) containing a predetermined quantity of antibodies recognized by said product.

24. A mono- or polyclonal antibody, its chimeric fragments or antibodies, characterized  
30 in that they are capable of specifically recognizing a product as defined in claim 20.

25. A method for the discriminatory detection of the presence of an antigen of *Mycobacterium tuberculosis* or *Mycobacterium africanum*, *Mycobacterium canetti*, *Mycobacterium microti*, *Mycobacterium bovis*, *Mycobacterium bovis* BCG, *Mycobacterium*



*tuberculosis* having the mutation CTG → CGG at codon 463 of gene katG and/or having no or very few IS6110 sequences inserted in their genome in a biological sample comprising the following steps:

- a) bringing the biological sample into contact with an antibody as claimed in claim 24,
- b) detecting the antigen-antibody complex formed.

---

26. A kit for the discriminatory detection of the presence of an antigen of *Mycobacterium tuberculosis* or *Mycobacterium africanum*, *Mycobacterium canetti*, *Mycobacterium microti*, *Mycobacterium bovis*, *Mycobacterium bovis BCG*, *Mycobacterium tuberculosis* having the mutation CTG → CGG at codon 463 of gene katG and/or having no or very few IS6110 sequences inserted in their genome, in a biological sample comprising the following steps:

- a) an antibody as claimed in claim 24,
- b) the reagents for constituting the medium suitable for the immunological reaction,
- c) the reagents allowing the detection of the antigen-antibody complexes produced by the immunological reaction.

27. An immunological composition, characterized in that it comprises at least one product as defined in claim 20.

28. A vaccine, characterized in that it comprises at least one product as defined in claim 20 in combination with a pharmaceutically compatible vehicle and, where appropriate, one or more appropriate immunity adjuvants.

29. An in vitro method for the detection and identification of *Mycobacterium tuberculosis* excepted *Mycobacterium tuberculosis* strain having the mutation CTG → CGG at codon 463 of gene katG and/or excepted *Mycobacterium tuberculosis* strain having no or very few IS6110 sequences inserted in their genome in a biological sample, comprising the following steps:

- a) isolation of the DNA from the biological sample to be analyzed or production of a cDNA from the RNA of the biological sample,

- b) detection of the nucleic acid sequences of the mycobacterium present in said biological sample,
- d) analysis for the presence or the absence of a nucleic acid fragment according to claims 2 or 3.

5

30. An *in vitro* method for the detection and identification of *Mycobacterium tuberculosis* excepted *Mycobacterium tuberculosis* strain having the mutation CTG → CGG at codon 463 of gene katG and/or excepted *Mycobacterium tuberculosis* strain having no or very few IS6110 sequences inserted in their genome in a biological sample, comprising the following steps:

- a) bringing the biological sample to be analyzed into contact with at least one pair of primers selected among nucleic acid fragments according to claims 1 to 4, 7 and 8, and more preferably selected among the primers chosen from the group comprising SEQ ID N°13, SEQ ID N°14, SEQ ID N°15, SEQ ID N°16, SEQ ID N°17, SEQ ID N°18, the DNA contained in the sample having been, where appropriate, made accessible to the hybridization beforehand,
- b) amplification of the DNA of the mycobacterium,
- c) visualization of the amplification of the DNA fragments.

20 31. A kit for the detection and identification of *Mycobacterium tuberculosis* excepted *Mycobacterium tuberculosis* strain having the mutation CTG → CGG at codon 463 of gene katG and/or excepted *Mycobacterium tuberculosis* strain having no or very few IS6110 sequences inserted in their genome in a biological sample, comprising the following elements:

- 25 a) at least one pair of primers selected among nucleic acid fragments according to claims 1 to 4, 7 and 8, and more preferably selected among the primers chosen from the group comprising SEQ ID N°13, SEQ ID N°14, SEQ ID N°15, SEQ ID N°16, SEQ ID N°17, SEQ ID N°18,
- b) the reagents necessary to carry out a DNA amplification reaction,
- 30 c) optionally, the necessary components which make it possible to verify or compare the sequence and/or the size of the amplified fragment.

32. A method for the detection *in vitro* of antibodies directed against *Mycobacterium tuberculosis* excepted *Mycobacterium Tuberculosis* having the mutation CTG → CGG at

codon 463 of gene KatG and/or having no or very few IS6110 sequences inserted in their genome, in a biological sample, comprising the following steps:

- a) bringing the biological sample into contact with at least one product as defined in claim 20,
- 5           b) detecting the antigen-antibody complex formed.

33. Use of TbD1 deletion as a as a genetic marker for the differentiation of Mycobacterium strain of Mycobacterium Tuberculosis complex.

10   34. Use of mmpL6<sup>551</sup> polymorphism as a genetic marker for the differentiation of Mycobacterium strain of Mycobacterium Tuberculosis complex.

35. Use of the geneic marker according to claims 33 and/or 34 in association with at least one genetic markers selected among RD1, RD2, RD3, RD4, RD5, RD6, RD7, RD8, RD9,  
15   RD10, Rd11, RD13, RD14, RvD1, RvD2, RvD3, RvD4, RvD5, katG<sup>463</sup>, gyrA<sup>95</sup>, oxyR<sup>285</sup>, pncA<sup>57</sup> for the differentiation of Mycobacterium strain of Mycobacterium Tuberculosis complex.

36. An *in vitro* method for the detection and identification of *Mycobacteria* from the  
20   *Mycobacterium Tuberculosis* complex in a biological sample, comprising the following steps:

- c) analysis for the presence or the absence of a nucleic acid fragment of the sequence TbD1 according to claims 2 or 3, and
- d) analysis of at least one additional genetic marker selected among RD1, RD2, RD3,  
25   RD4, RD5, RD6, RD7, RD8, RD9, RD10, Rd11, RD13, RD14, RvD1, RvD2, RvD3, RvD4, RvD5, katG<sup>463</sup>, gyrA<sup>95</sup>, oxyR<sup>285</sup>, pncA<sup>57</sup>.

37. The *in vitro* method of claim 36 wherein two additional markers are used, preferably RD4 and RD9.

30

38. The method according to claim 36 wherein the analysis is performed by a technique selected among sequence hybridization, nucleic acid amplification, antigen-antibody complex.

39. A kit for the detection and identification of *Mycobacteria* from the *Mycobacterium Tuberculosis* complex in a biological sample comprising the following elements:

- e) at least one pair of primers selected among nucleic acid fragments according to claims 1 to 4, 7 and 8, and more preferably selected among the primers chosen from the group comprising SEQ ID N°13, SEQ ID N°14, SEQ ID N°15, SEQ ID N°16, SEQ ID N°17, SEQ ID N°18,
- f) at least one pair of primers specific of the genetic markers selected among RD1, RD2, RD3, RD4, RD5, RD6, RD7, RD8, RD9, RD10, Rd11, RD13, RD14, RvD1, RvD2, RvD3, RvD4, RvD5, katG<sup>463</sup>, gyrA<sup>95</sup>, oxyR<sup>285</sup>, pncA<sup>57</sup>.
- g) the reagents necessary to carry out a DNA amplification reaction,
- h) optionally, the necessary components which make it possible to verify or compare the sequence and/or the size of the amplified fragment.

40. A kit according to claim 39 comprising the following elements:

- f) at least one pair of primers selected among nucleic acid fragments according to claims 1 to 4, 7 and 8, and more preferably selected among the primers chosen from the group comprising SEQ ID N°13, SEQ ID N°14, SEQ ID N°15, SEQ ID N°16, SEQ ID N°17, SEQ ID N°18,
- g) one pair of primers specific of the genetic markers RD4,
- h) one pair of primers specific of the genetic markers RD9,
- i) the reagents necessary to carry out a DNA amplification reaction,
- j) optionally, the necessary components which make it possible to verify or compare the sequence and/or the size of the amplified fragment.

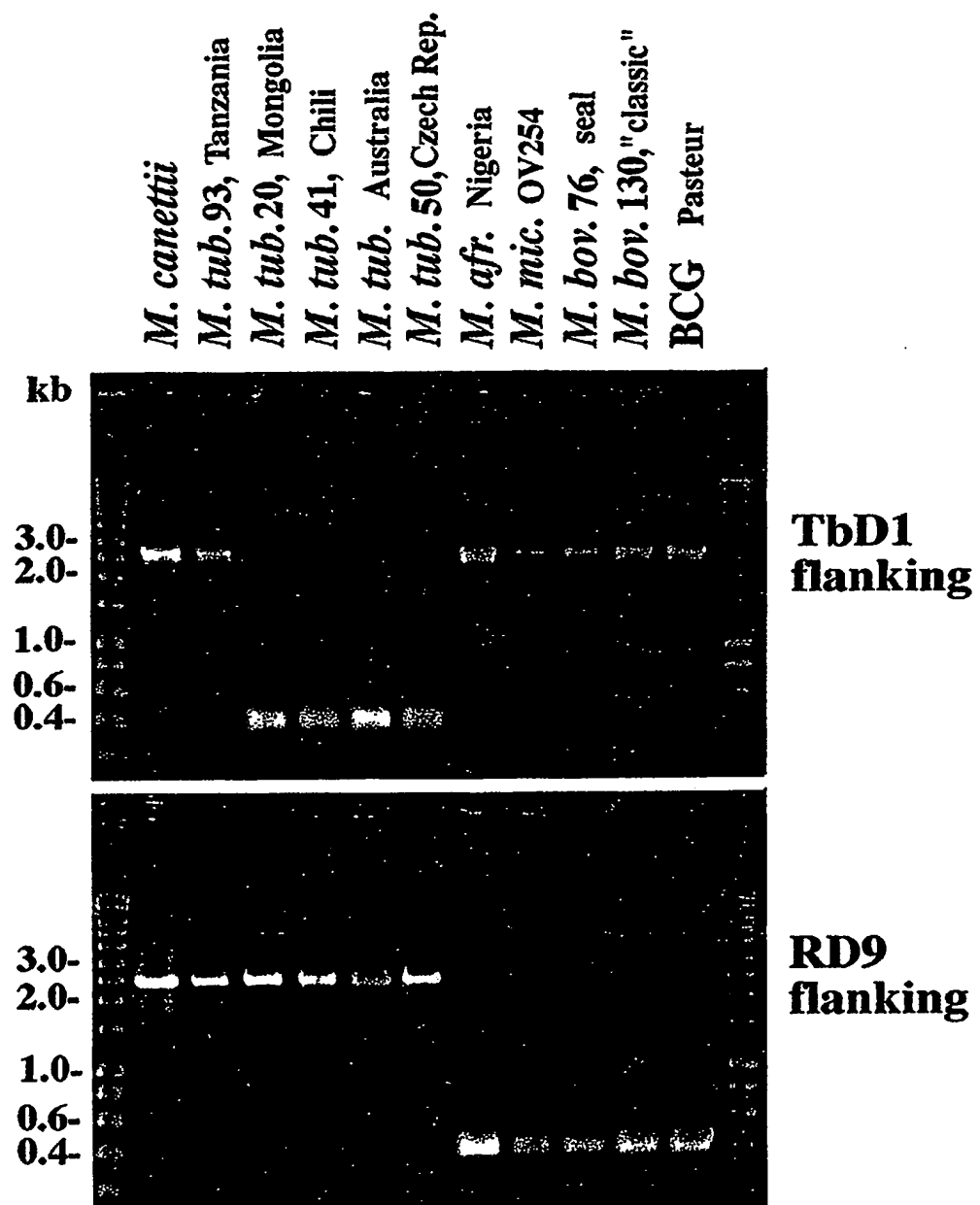
# PATENT APPLICATION

**TITLE:** DELETED SEQUENCE IN *M. TUBERCULOSIS*, METHOD FOR  
5 DETECTING MYCOBACTERIA USING THESE SEQUENCES AND  
VACCINES

**APPLICANTS:** INSTITUT PASTEUR

## **ABSTRACT:**

15 The present invention is the identification of a nucleotide sequence which make it possible in particular to distinguish an infection resulting from *Mycobacterium tuberculosis* from an infection resulting from *Mycobacterium africanum*, *Mycobacterium canetti*, *Mycobacterium microti*, *Mycobacterium bovis*, *Mycobacterium bovis BCG*. The subject of  
20 the present invention is also a method for detecting the sequences in question by the products of expression of these sequences and the kits for carrying out these methods. Finally, the subject of the present invention is novel vaccines.

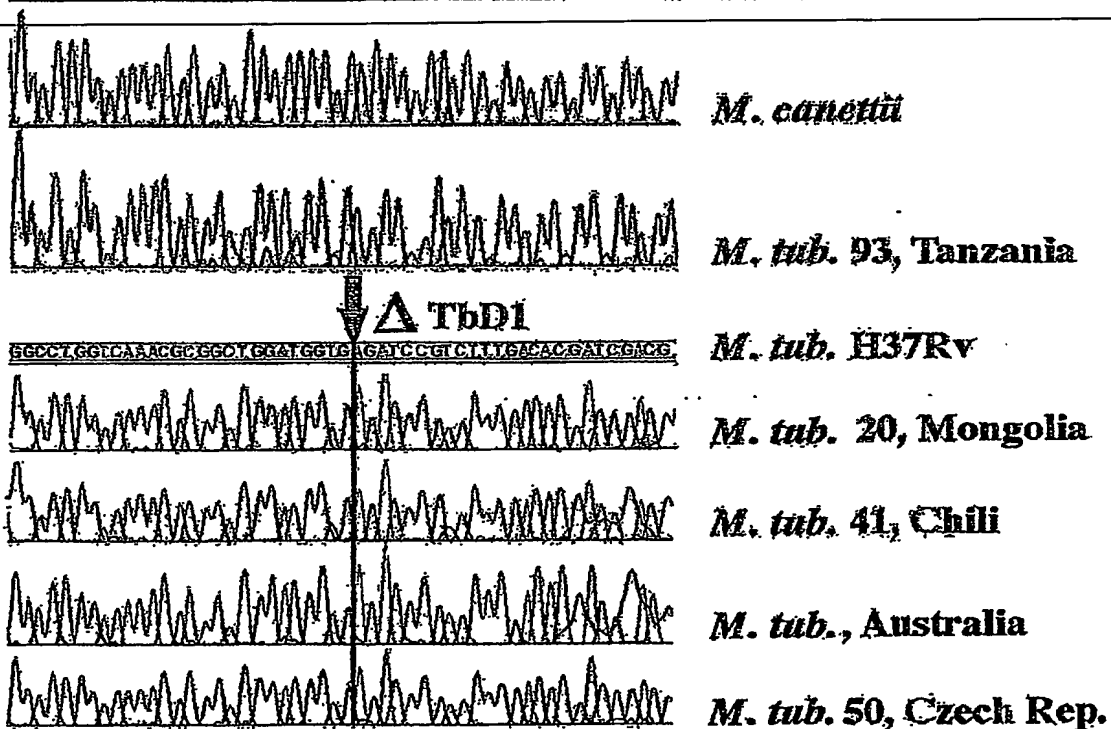


**Figure 1**

### TbD1-region

GGCTGCTOAAAGCGCGCTGGTGGTGGTGGTGGCGGTGGCGGTGGTGGCGG

***M. bovis* AF2122/97**



## Figure 2

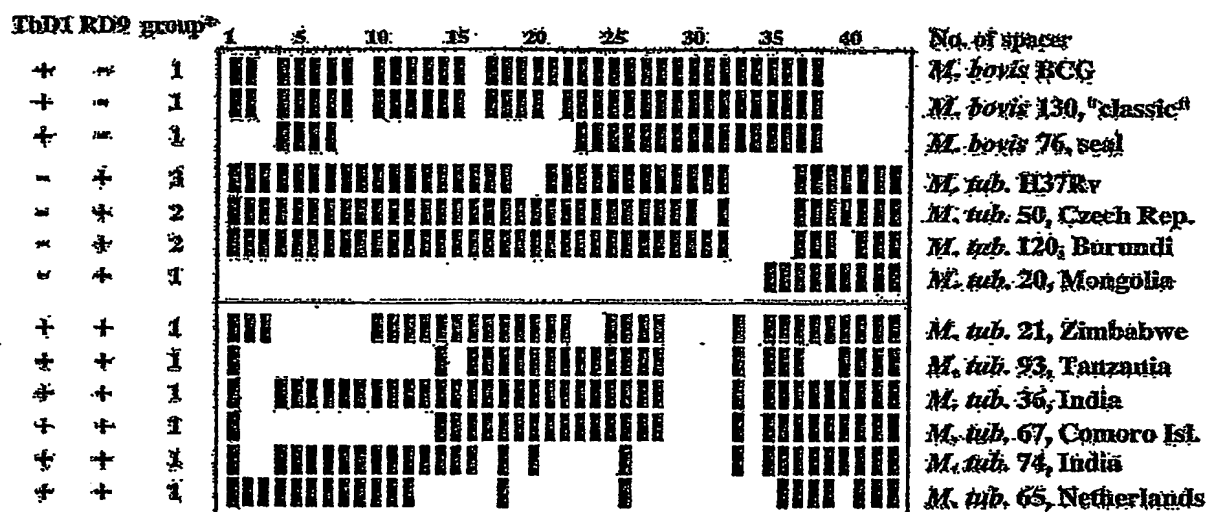


Fig. 1c: Spoligotypes and presence/absence of TbD1 and RD9 regions of selected *M. tuberculosis* and *M. bovis* strains. Numbers correspond to strain designation used in Ref. 8 and 9.

Figure 3



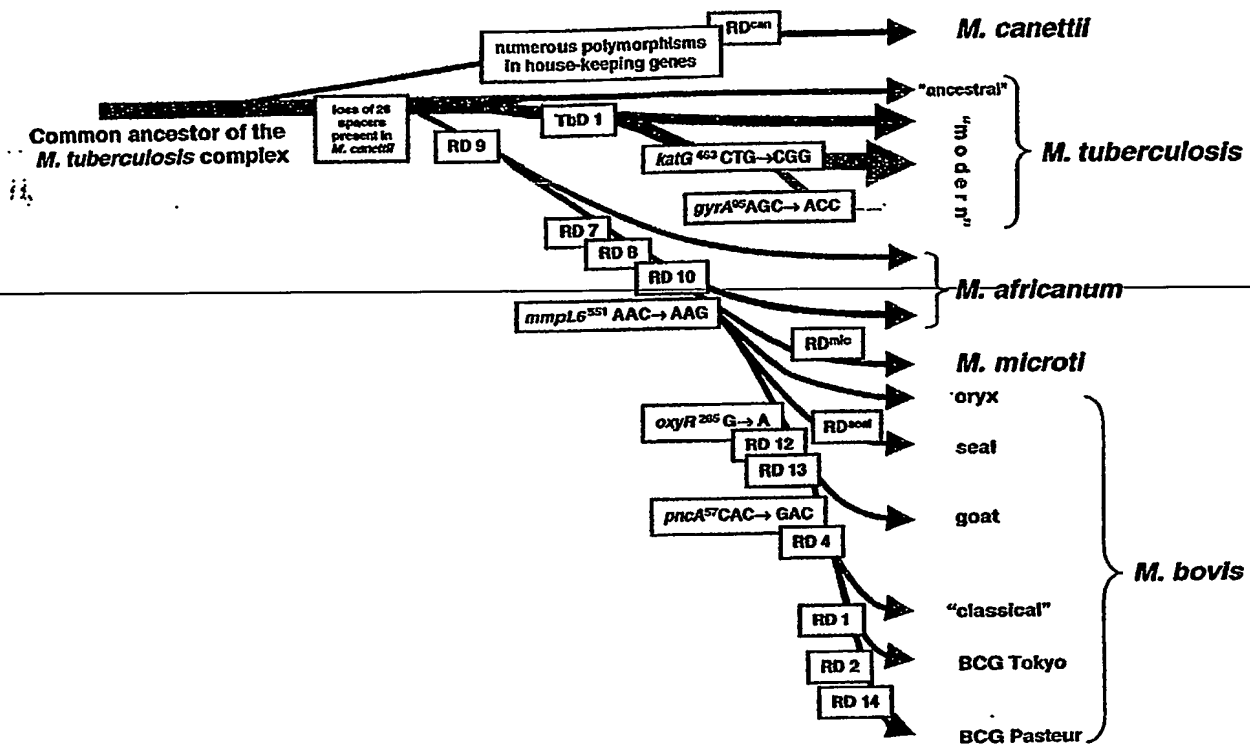


Figure 4

## SEQUENCE LISTING

&lt;110&gt; Institut PASTEUR

<120> DELETED SEQUENCE IN M. TUBERCULOSIS, METHOD FOR  
DETECTING MYCOBACTERIA USING THESE SEQUENCES AND  
VACCINES

&lt;130&gt; D20110

&lt;160&gt; 20

&lt;170&gt; PatentIn Ver. 2.1

&lt;210&gt; 1

&lt;211&gt; 3953

&lt;212&gt; DNA

&lt;213&gt; Mycobacterium tuberculosis

&lt;220&gt;

&lt;221&gt; CDS

&lt;222&gt; (735) .. (3638)

&lt;400&gt; 1

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tccagcgcg ccatcagcga tgaactctgg gacctgctac cgggtacct catcttccgg 60
tccatcatcc ccaaccggcc gccacccag gacacgggtg aagccctcgt cgacgacgtg 120
atactcccca gcctcaccgg atccaccggt tgagtcagcg gtgcgaatgg ctgggcaccg 180
ttgtgggtgc cgggtccgta cgtactgtt gaatccgcgg atccccgcct gaggtacggg 240
gcgtgggtgc gccccgggca atagcgtcgc cggttatcga aaggctaacg ggtgcagggg 300
atttcagtga ctggcctggt caaacgcggc tggatgggtg tggttgccgt ggcgggtggtg 360
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His Thr Ile Arg Arg Leu Ser Leu Pro Ile Leu Leu Phe Trp Val Gly
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Met	Leu	Leu	Phe	Val	Tyr	Arg	Ser	Val	Val	Thr	Met	Val	Leu	Val	Leu		
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ctc	gga	aac	gcc	ggg	gta	atc	ggg	ctg	tcg	aca	tac	tcg	acg	aat	ctg	1490	
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Ala	Phe	Tyr	Thr	Met	Tyr	Arg	Gly	Thr	Ala	His	Val	Val	Leu	Gly	Ser		

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Val Ala Gln Ser Ser Pro Asp Asp Pro Ser Leu Gln Ala Met Lys Arg  
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 Asp Thr Leu Leu Arg Asn Leu Ser Asn Asp Thr Lys His Val Glu His  
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 Ala His Thr Pro Pro Pro Ala Gly Val Lys Ala Tyr Val Thr Gly Ala  
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 Lys Val Thr Gly Ile Thr Leu Val Val Ile Ala Val Met Leu Leu Phe  
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 Val Tyr Arg Ser Val Val Thr Met Val Leu Val Leu Ile Thr Val Leu  
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 Gly Val Ile Gly Leu Ser Thr Tyr Ser Thr Asn Leu Leu Thr Leu Leu  
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Ser	Ser	Ile	Pro	Phe	Gln	Ile	Ser	Met	Gln	Ser	Val	Gly	Gln	Ile	Gln
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Cys	Tyr	Asp	Ile	Pro	Ser	Cys	Trp	Ala	Leu	Arg	Ser	Val	Phe	Asp	Thr
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Thr	Leu	Asp	Lys	Leu	Ala	Ala	Ile	Gln	Pro	Gln	Leu	Val	Ala	Leu	Leu	610	615	620	
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Glu	Asn	Ala	Ala	Ala	Met	Gly	Gln	Ala	Phe	Asp	Ala	Ala	Lys	Asn	Asp	660	665	670	
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Arg	Gly	Leu	Lys	Leu	Phe	Leu	Ser	Ala	Asp	Gly	Lys	Ala	Ala	Arg	Met	690	695	700	
Ile	Ile	Ser	His	Glu	Gly	Asp	Pro	Ala	Thr	Pro	Glu	Gly	Ile	Ser	His	705	710	715	720
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Ser	Phe	Gly	Leu	Ser	Val	Leu	Val	Trp	Gln	His	Leu	Leu	Gly	Ile	Gln	805	810	815	
Leu	Tyr	Trp	Ile	Val	Leu	Ala	Leu	Ala	Val	Ile	Leu	Leu	Leu	Ala	Val	820	825	830	
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Pro	Ser	Ile	Ala	Val	Leu	Leu	Gly	Arg	Trp	Phe	Trp	Trp	Pro	Gln	Arg	915	920	925	

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Val Ala Asn Asp Ile Lys Pro Phe Asn Pro Lys Gln Val Thr Leu Glu  
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Val Phe Gly Ala Pro Gly Thr Val Ala Thr Ile Asn Tyr Leu Asp Val  
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Asp Ala Thr Pro Arg Gln Val Leu Asp Thr Thr Leu Pro Trp Ser Tyr  
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Thr Ile Thr Thr Thr Leu Pro Ala Val Phe Ala Asn Val Val Ala Gln  
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Gly Asp Ser Asn Ser Ile Gly Cys Arg Ile Thr Val Asn Gly Val Val  
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Arg Leu Ser Leu Pro Ile Leu Leu Phe Trp Val Gly Val Ala Ala Ile
20 25 30
acc aat gcc gcc gtg ccg caa ttg gag gtg gtc ggg gag gcg cat aac 144
Thr Asn Ala Ala Val Pro Gln Leu Glu Val Val Gly Glu Ala His Asn
35 40 45
gtc gca cag agc tcc ccg gat gac ccg tcg ctg cag gcg atg aaa cgc 192
Val Ala Gln Ser Ser Pro Asp Asp Pro Ser Leu Gln Ala Met Lys Arg

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gac acc ctg ctc cgc aac ctt tca aac gac acc aaa cac gtc gag cac Asp Thr Leu Leu Arg Asn Leu Ser Asn Asp Thr Lys His Val Glu His 100 105 110	336		
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Ala Gly Ala Val Tyr Cys Leu Ser Phe Thr Arg Leu Pro Tyr Phe Gln			
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Ser Leu Gly Ile Pro Ala Ser Ile Gly Val Met Ile Ala Leu Ala Ala			
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Cys Phe Glu Pro Lys Arg Arg Met Arg Thr Arg Gly Trp Arg Arg Ile			
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Gly Thr Ala Ile Val Arg Trp Pro Gly Pro Ile Leu Ala Val Ala Cys			
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Tyr Asp Ala Arg Tyr Tyr Met Pro Ala Thr Ala Pro Ala Asn Ile Gly			
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Leu Leu Met Ile Glu Thr Asp His Asp Met Arg Asn Pro Ala Asp Met			
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ctc atc ttg gat agg atc gcc aag gct gtc ttc cat ctg ccc ggc ata			1392
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Asn Leu Lys Tyr Gln Arg Asp Arg Ala Ala Asp Leu Leu Lys Gln Ala			
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Glu Glu Leu Gly Lys Thr Ile Glu Ile Leu Gln Arg Gln Tyr Ala Leu			
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&lt;211&gt; 967

&lt;212&gt; PRT

&lt;213&gt; Mycobacterium tuberculosis

&lt;220&gt;

&lt;223&gt; mmpL6 sequence and protein

&lt;400&gt; 6

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Thr	Asn	Ala	Ala	Val	Pro	Gln	Leu	Glu	Val	Val	Gly	Glu	Ala	His	Asn	35	40	45	
Val	Ala	Gln	Ser	Ser	Pro	Asp	Asp	Pro	Ser	Leu	Gln	Ala	Met	Lys	Arg	50	55	60	
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Gly	Glu	Ala	Leu	Ser	Ile	Glu	Ser	Val	Asp	Ala	Val	Arg	Asp	Ile	Val	145	150	155	160
Ala	His	Thr	Pro	Pro	Pro	Ala	Gly	Val	Lys	Ala	Tyr	Val	Thr	Gly	Ala	165	170	175	
Ala	Pro	Leu	Met	Ala	Asp	Gln	Phe	Gln	Val	Gly	Ser	Lys	Gly	Thr	Ala	180	185	190	
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Ser	Leu	Gly	Ile	Pro	Ala	Ser	Ile	Gly	Val	Met	Ile	Ala	Leu	Ala	Ala				



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Tyr	Met	Ala	Ala	Glu	Arg	His	Phe	Pro	Gln	Ala	Arg	Leu	Asn	Pro	Glu
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Leu	Ile	Leu	Asp	Arg	Ile	Ala	Lys	Ala	Val	Phe	His	Leu	Pro	Gly	Ile
	450					455					460				
Gly	Leu	Val	Gln	Ala	Met	Thr	Arg	Pro	Leu	Gly	Thr	Pro	Ile	Asp	His
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Glu	Glu	Leu	Gly	Lys	Thr	Ile	Glu	Ile	Leu	Gln	Arg	Gln	Tyr	Ala	Leu
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Gln	Gln	Glu	Leu	Ala	Ala	Ala	Thr	His	Glu	Gln	Ala	Glu	Ser	Phe	His
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Gln	Thr	Ile	Ala	Thr	Val	Lys	Glu	Leu	Arg	Asp	Arg	Ile	Ala	Asn	Phe
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Cys	Tyr	Asp	Ile	Pro	Ser	Cys	Trp	Ala	Leu	Arg	Ser	Val	Phe	Asp	Thr
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Ile	Asp	Gly	Ile	Asp	Gln	Leu	Gly	Glu	Gln	Leu	Ala	Ser	Val	Thr	Val
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	610					615					620				
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 Asp Ser Phe Tyr Leu Pro Pro Glu Ala Phe Asp Asn Pro Asp Phe Gln  
 675 680 685  
 Arg Gly Leu Lys Leu Phe Leu Ser Ala Asp Gly Lys Ala Ala Arg Met  
 690 695 700  
 Ile Ile Ser His Glu Gly Asp Pro Ala Thr Pro Glu Gly Ile Ser His  
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 725 730 735  
 Met Ala Gly Ala Gly Ile Tyr Leu Ala Gly Thr Ala Ala Thr Phe Lys  
 740 745 750  
 Asp Ile Gln Asp Gly Ala Thr Tyr Asp Leu Leu Ile Ala Gly Ile Ala  
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Arg Leu Ser Leu Pro Ile Leu Leu Phe Trp Val Gly Val Ala Ala Ile	
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acc aat gcc gcc gtg ccg caa ttg gag gtg gtc ggg gag gcg cat aac	144
Thr Asn Ala Ala Val Pro Gln Leu Glu Val Val Gly Glu Ala His Asn	
35 40 45	
gtc gca cag agc tcc ccg gat gac ccg tcg ctg cag gcg atg aaa cgc	192
Val Ala Gln Ser Ser Pro Asp Asp Pro Ser Leu Gln Ala Met Lys Arg	
50 55 60	
atc ggc aag gtg ttc cac gag ttc gat tcc gac agt gcg gcc atg atc	240
Ile Gly Lys Val Phe His Glu Phe Asp Ser Asp Ser Ala Ala Met Ile	
65 70 75 80	
gtc ttg gaa ggc gat aag ccg ctc ggc aac gac gcc cac cgg ttc tac	288
Val Leu Glu Gly Asp Lys Pro Leu Gly Asn Asp Ala His Arg Phe Tyr	
85 90 95	
gac acc ctg ctc cgc aac ctt tca aac gac acc aaa cac gtc gag cac	336
Asp Thr Leu Leu Arg Asn Leu Ser Asn Asp Thr Lys His Val Glu His	
100 105 110	
gtt cag gac ttc tgg ggc gat ccg ctg acc gcg gcc ggc tcg caa agc	384
Val Gln Asp Phe Trp Gly Asp Pro Leu Thr Ala Ala Gly Ser Gln Ser	
115 120 125	
acc gac ggc aaa gcc gcc tac gtt cag gtc tat ctc gcc ggc aac caa	432
Thr Asp Gly Lys Ala Ala Tyr Val Gln Val Tyr Leu Ala Gly Asn Gln	
130 135 140	
ggc gag gcg ttg tca atc gag tcc gtc gac gcg gtg cgc gac atc gtc	480
Gly Glu Ala Leu Ser Ile Glu Ser Val Asp Ala Val Arg Asp Ile Val	
145 150 155 160	
gcc cat acg cca cca ccg gcc ggg gtc aag gcc tac gtc acc ggc gcg	528
Ala His Thr Pro Pro Pro Ala Gly Val Lys Ala Tyr Val Thr Gly Ala	
165 170 175	
gcc ccg ctc atg gcc gat cag ttt cag gtg ggc agc aaa gga acc gcg	576

Ala	Pro	Leu	Met	Ala	Asp	Gln	Phe	Gln	Val	Gly	Ser	Lys	Gly	Thr	Ala	
			180					185					190			
aaa	gtt	acc	ggg	ata	act	ctg	gtt	gtg	atc	gcg	gtg	atg	ttg	ctc	ttc	624
Lys	Val	Thr	Gly	Ile	Thr	Leu	Val	Val	Ile	Ala	Val	Met	Leu	Leu	Phe	
		195					200					205				
gta	tac	cgt	tcc	gtc	gtc	acc	atg	gtc	ctg	gtg	ctt	atc	acg	gtt	ctt	672
Val	Tyr	Arg	Ser	Val	Val	Thr	Met	Val	Leu	Val	Leu	Ile	Thr	Val	Leu	
	210					215					220					
att	gag	ttg	gcc	gcg	gcc	cgc	ggg	atc	gtc	gct	ttt	ctc	gga	aac	gcc	720
Ile	Glu	Leu	Ala	Ala	Ala	Arg	Gly	Ile	Val	Ala	Phe	Leu	Gly	Asn	Ala	
225					230					235					240	
ggg	gta	atc	ggg	ctg	tcg	aca	tac	tcg	acg	aat	ctg	ctc	aca	cta	ttg	768
Gly	Val	Ile	Gly	Leu	Ser	Thr	Tyr	Ser	Thr	Asn	Leu	Leu	Thr	Leu	Leu	
				245					250					255		
gta	atc	gcg	gcg	ggc	aca	gac	tac	gcg	att	ttt	gtc	ctc	ggc	cgc	tat	816
Val	Ile	Ala	Ala	Gly	Thr	Asp	Tyr	Ala	Ile	Phe	Val	Leu	Gly	Arg	Tyr	
			260					265					270			
cac	gag	gcg	cgc	tac	gcc	gca	cag	gat	cgg	gaa	acg	gcc	ttc	tac	acg	864
His	Glu	Ala	Arg	Tyr	Ala	Ala	Gln	Asp	Arg	Glu	Thr	Ala	Phe	Tyr	Thr	
		275					280					285				
atg	tat	cgc	ggg	acc	gcc	cac	gtc	gtc	ttg	ggc	tcg	ggg	ctg	acc	gtt	912
Met	Tyr	Arg	Gly	Thr	Ala	His	Val	Val	Leu	Gly	Ser	Gly	Leu	Thr	Val	
	290					295					300					
gcc	ggc	gcg	gtg	tat	tgc	ctg	agc	ttt	acc	cgg	cta	ccc	tat	ttt	caa	960
Ala	Gly	Ala	Val	Tyr	Cys	Leu	Ser	Phe	Thr	Arg	Leu	Pro	Tyr	Phe	Gln	
305					310					315					320	
agc	ctg	ggg	att	ccc	gcc	tcg	ata	ggg	gtg	atg	att	gcg	ttg	gca	gcc	1008
Ser	Leu	Gly	Ile	Pro	Ala	Ser	Ile	Gly	Val	Met	Ile	Ala	Leu	Ala	Ala	
				325					330					335		
gcg	ctc	agc	ctg	gcc	cca	tcc	gtg	ctc	atc	ttg	ggc	agt	cgt	ttc	ggg	1056
Ala	Leu	Ser	Leu	Ala	Pro	Ser	Val	Leu	Ile	Leu	Gly	Ser	Arg	Phe	Gly	
				340				345					350			
tgt	ttc	gaa	ccc	aag	cgc	agg	atg	agg	acc	agg	gga	tgg	cgg	cgc	atc	1104
Cys	Phe	Glu	Pro	Lys	Arg	Arg	Met	Arg	Thr	Arg	Gly	Trp	Arg	Arg	Ile	
		355					360					365				
ggc	acg	gcc	atc	gtg	cgt	tgg	ccg	gga	ccc	atc	ctg	gca	gtg	gcg	tgc	1152
Gly	Thr	Ala	Ile	Val	Arg	Trp	Pro	Gly	Pro	Ile	Leu	Ala	Val	Ala	Cys	
	370					375					380					
gca	att	gcg	gtg	gtg	ggg	ctg	ctc	gcg	ctg	ccg	gga	tac	aaa	acg	agc	1200
Ala	Ile	Ala	Val	Val	Gly	Leu	Leu	Ala	Leu	Pro	Gly	Tyr	Lys	Thr	Ser	
385					390					395					400	
tac	gac	gct	cgc	tat	tac	atg	ccc	gcc	acc	gcc	ccg	gcc	aat	att	ggc	1248
Tyr	Asp	Ala	Arg	Tyr	Tyr	Met	Pro	Ala	Thr	Ala	Pro	Ala	Asn	Ile	Gly	
				405					410					415		
tac	atg	gcc	gcg	gag	cga	cat	ttt	ccc	caa	gcg	cgg	ctg	aat	ccc	gaa	1296
Tyr	Met	Ala	Ala	Glu	Arg	His	Phe	Pro	Gln	Ala	Arg	Leu	Asn	Pro	Glu	

420	425	430	
cta ctg atg atc gag acg gat cac gat atg cgc aat ccg gcc gac atg			1344
Leu Leu Met Ile Glu Thr Asp His Asp Met Arg Asn Pro Ala Asp Met			
435	440	445	
ctc atc ttg gat agg atc gcc aag gct gtc ttc cat ctg ccc ggc ata			1392
Leu Ile Leu Asp Arg Ile Ala Lys Ala Val Phe His Leu Pro Gly Ile			
450	455	460	
ggg ctg gtg cag gcc atg acc cgg ccg cta gga acc ccg att gac cac			1440
Gly Leu Val Gln Ala Met Thr Arg Pro Leu Gly Thr Pro Ile Asp His			
465	470	475	480
agc tcg ata ccg ttt cag atc agc atg caa agc gtc ggc cag att cag			1488
Ser Ser Ile Pro Phe Gln Ile Ser Met Gln Ser Val Gly Gln Ile Gln			
485	490	495	
<hr/>			
aat ctc aag tat cag agg gac cga gca gcc gac ttg ctg aag cag gcc			1536
Asn Leu Lys Tyr Gln Arg Asp Arg Ala Ala Asp Leu Leu Lys Gln Ala			
500	505	510	
gaa gag ctg ggg aag acg atc gaa atc ttg cag cgc caa tat gcc cta			1584
Glu Glu Leu Gly Lys Thr Ile Glu Ile Leu Gln Arg Gln Tyr Ala Leu			
515	520	525	
cag cag gaa ctc gcg gcc gct act cac gag caa gcc gaa agc ttt cac			1632
Gln Gln Glu Leu Ala Ala Ala Thr His Glu Gln Ala Glu Ser Phe His			
530	535	540	
caa acg atc gcc acg gta aag gaa ctg cga gat agg atc gcc aat ttc			1680
Gln Thr Ile Ala Thr Val Lys Glu Leu Arg Asp Arg Ile Ala Asn Phe			
545	550	555	560
gac gat ttc ttc agg ccg att cgt agt tac ttt tac tgg gaa aag cac			1728
Asp Asp Phe Phe Arg Pro Ile Arg Ser Tyr Phe Tyr Trp Glu Lys His			
565	570	575	
tgc tac gat atc ccg agc tgc tgg gcg ctg			1758
Cys Tyr Asp Ile Pro Ser Cys Trp Ala Leu			
580	585		

&lt;210&gt; 8

&lt;211&gt; 586

&lt;212&gt; PRT

&lt;213&gt; Mycobacterium tuberculosis

&lt;220&gt;

&lt;223&gt; mmpL6 truncated sequence and protein

&lt;400&gt; 8

Met	Ser	Asn	His	His	Arg	Pro	Arg	Pro	Trp	Leu	Pro	His	Thr	Ile	Arg
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Arg	Leu	Ser	Leu	Pro	Ile	Leu	Leu	Phe	Trp	Val	Gly	Val	Ala	Ala	Ile
	20						25						30		

Thr	Asn	Ala	Ala	Val	Pro	Gln	Leu	Glu	Val	Val	Gly	Glu	Ala	His	Asn
	35					40						45			

Val Ala Gln Ser Ser Pro Asp Asp Pro Ser Leu Gln Ala Met Lys Arg  
 50 55 60  
 Ile Gly Lys Val Phe His Glu Phe Asp Ser Asp Ser Ala Ala Met Ile  
 65 70 75 80  
 Val Leu Glu Gly Asp Lys Pro Leu Gly Asn Asp Ala His Arg Phe Tyr  
 85 90 95  
 Asp Thr Leu Leu Arg Asn Leu Ser Asn Asp Thr Lys His Val Glu His  
 100 105 110  
 Val Gln Asp Phe Trp Gly Asp Pro Leu Thr Ala Ala Gly Ser Gln Ser  
 115 120 125  
 Thr Asp Gly Lys Ala Ala Tyr Val Gln Val Tyr Leu Ala Gly Asn Gln  
 130 135 140  
 Gly Glu Ala Leu Ser Ile Glu Ser Val Asp Ala Val Arg Asp Ile Val  
 145 150 155 160  
 Ala His Thr Pro Pro Pro Ala Gly Val Lys Ala Tyr Val Thr Gly Ala  
 165 170 175  
 Ala Pro Leu Met Ala Asp Gln Phe Gln Val Gly Ser Lys Gly Thr Ala  
 180 185 190  
 Lys Val Thr Gly Ile Thr Leu Val Val Ile Ala Val Met Leu Leu Phe  
 195 200 205  
 Val Tyr Arg Ser Val Val Thr Met Val Leu Val Leu Ile Thr Val Leu  
 210 215 220  
 Ile Glu Leu Ala Ala Ala Arg Gly Ile Val Ala Phe Leu Gly Asn Ala  
 225 230 235 240  
 Gly Val Ile Gly Leu Ser Thr Tyr Ser Thr Asn Leu Leu Thr Leu Leu  
 245 250 255  
 Val Ile Ala Ala Gly Thr Asp Tyr Ala Ile Phe Val Leu Gly Arg Tyr  
 260 265 270  
 His Glu Ala Arg Tyr Ala Ala Gln Asp Arg Glu Thr Ala Phe Tyr Thr  
 275 280 285  
 Met Tyr Arg Gly Thr Ala His Val Val Leu Gly Ser Gly Leu Thr Val  
 290 295 300  
 Ala Gly Ala Val Tyr Cys Leu Ser Phe Thr Arg Leu Pro Tyr Phe Gln  
 305 310 315 320  
 Ser Leu Gly Ile Pro Ala Ser Ile Gly Val Met Ile Ala Leu Ala Ala  
 325 330 335  
 Ala Leu Ser Leu Ala Pro Ser Val Leu Ile Leu Gly Ser Arg Phe Gly  
 340 345 350  
 Cys Phe Glu Pro Lys Arg Arg Met Arg Thr Arg Gly Trp Arg Arg Ile  
 355 360 365  
 Gly Thr Ala Ile Val Arg Trp Pro Gly Pro Ile Leu Ala Val Ala Cys

370 375 380

Ala Ile Ala Val Val Gly Leu Leu Ala Leu Pro Gly Tyr Lys Thr Ser  
385 390 395 400

Tyr Asp Ala Arg Tyr Tyr Met Pro Ala Thr Ala Pro Ala Asn Ile Gly  
405 410 415

Tyr Met Ala Ala Glu Arg His Phe Pro Gln Ala Arg Leu Asn Pro Glu  
420 425 430

Leu Leu Met Ile Glu Thr Asp His Asp Met Arg Asn Pro Ala Asp Met  
435 440 445

Leu Ile Leu Asp Arg Ile Ala Lys Ala Val Phe His Leu Pro Gly Ile  
450 455 460

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Gly Leu Val Gln Ala Met Thr Arg Pro Leu Gly Thr Pro Ile Asp His  
465 470 475 480

Ser Ser Ile Pro Phe Gln Ile Ser Met Gln Ser Val Gly Gln Ile Gln  
485 490 495

Asn Leu Lys Tyr Gln Arg Asp Arg Ala Ala Asp Leu Leu Lys Gln Ala  
500 505 510

Glu Glu Leu Gly Lys Thr Ile Glu Ile Leu Gln Arg Gln Tyr Ala Leu  
515 520 525

Gln Gln Glu Leu Ala Ala Ala Thr His Glu Gln Ala Glu Ser Phe His  
530 535 540

Gln Thr Ile Ala Thr Val Lys Glu Leu Arg Asp Arg Ile Ala Asn Phe  
545 550 555 560

Asp Asp Phe Phe Arg Pro Ile Arg Ser Tyr Phe Tyr Trp Glu Lys His  
565 570 575

Cys Tyr Asp Ile Pro Ser Cys Trp Ala Leu  
580 585

<210> 9  
<211> 447  
<212> DNA  
<213> Mycobacterium tuberculosis

<220>  
<221> CDS  
<222> (1)..(444)

<220>  
<223> mmpS6 sequence and protein

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Val Gln Gly Ile Ser Val Thr Gly Leu Val Lys Arg Gly Trp Met Val  
1 5 10 15  
ctg gtt gcc gtg gcg gtg gtg gcg gtc gcg gga ttc agc gtc tat cgg 96

Leu	Val	Ala	Val	Ala	Val	Val	Ala	Val	Ala	Gly	Phe	Ser	Val	Tyr	Arg	
						20		25		30						
ttg	cac	ggc	atc	ttc	ggc	tcg	cac	gac	acc	acc	tcg	acc	gcc	ggg	ggg	
Leu	His	Gly	Ile	Phe	Gly	Ser	His	Asp	Thr	Thr	Ser	Thr	Ala	Gly	Gly	
						40		45								
gtc	gcg	aac	gac	atc	aag	ccg	ttc	aac	ccc	aaa	cag	gta	acc	ctc	gag	
Val	Ala	Asn	Asp	Ile	Lys	Pro	Phe	Asn	Pro	Lys	Gln	Val	Thr	Leu	Glu	
						55		60								
gtc	ttt	ggc	gct	ccc	gga	acc	gtg	gca	acg	atc	aat	tat	ctg	gac	gtg	
Val	Phe	Gly	Ala	Pro	Gly	Thr	Val	Ala	Thr	Ile	Asn	Tyr	Leu	Asp	Val	
						70		75								
						80										
gat	gcc	aca	cct	cgg	caa	gtc	ctg	gac	acg	acc	ctg	ccg	tgg	tca	tac	
Asp	Ala	Thr	Pro	Arg	Gln	Val	Leu	Asp	Thr	Thr	Leu	Pro	Trp	Ser	Tyr	
						85		90								
						95										
acg	atc	acg	acg	acc	ctg	ccc	gcg	gtc	ttc	gcc	aat	gtt	gtc	gcg	caa	
Thr	Ile	Thr	Thr	Thr	Leu	Pro	Ala	Val	Phe	Ala	Asn	Val	Val	Ala	Gln	
						100		105								
						110										
ggc	gac	agc	aat	tcc	atc	ggc	tgc	cgc	atc	acc	gtc	aac	ggg	gta	gtc	
Gly	Asp	Ser	Asn	Ser	Ile	Gly	Cys	Arg	Ile	Thr	Val	Asn	Gly	Val	Val	
						115		120								
						125										
aag	gac	gaa	agg	atc	gtc	aac	gaa	gtg	cgc	gcc	tat	acc	ttc	tgc	ctc	
Lys	Asp	Glu	Arg	Ile	Val	Asn	Glu	Val	Arg	Ala	Tyr	Thr	Phe	Cys	Leu	
						130		135								
						140										
gac	aag	tcc	tca	tga												
Asp	Lys	Ser	Ser													
						145										
						147										

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<210> 10
<211> 148
<212> PRT
<213> Mycobacterium tuberculosis
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<220>  
<223> mmpS6 sequence and protein

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Leu Val Ala Val Ala Val Val Ala Val Ala Gly Phe Ser Val Tyr Arg
      20          25          30

Leu His Gly Ile Phe Gly Ser His Asp Thr Thr Ser Thr Ala Gly Gly
      35          40          45

Val Ala Asn Asp Ile Lys Pro Phe Asn Pro Lys Gln Val Thr Leu Glu
      50          55          60

Val Phe Gly Ala Pro Gly Thr Val Ala Thr Ile Asn Tyr Leu Asp Val
      65          70          75          80

Asp Ala Thr Pro Arg Gln Val Leu Asp Thr Thr Leu Pro Trp Ser Tyr

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	85		90		95
Thr Ile Thr Thr Thr Leu Pro Ala Val Phe Ala Asn Val Val Ala Gln					
	100		105		110
Gly Asp Ser Asn Ser Ile Gly Cys Arg Ile Thr Val Asn Gly Val Val					
	115		120		125
Lys Asp Glu Arg Ile Val Asn Glu Val Arg Ala Tyr Thr Phe Cys Leu					
	130		135		140
Asp Lys Ser Ser					
145					
:					

&lt;210&gt; 11

&lt;211&gt; 399

&lt;212&gt; DNA

&lt;213&gt; Mycobacterium tuberculosis

&lt;220&gt;

&lt;221&gt; CDS

&lt;222&gt; (1) .. (399)

&lt;220&gt;

&lt;223&gt; mmpS6 truncated sequence and protein

&lt;400&gt; 11

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Leu Val Ala Val Ala Val Val Ala Val Ala Gly Phe Ser Val Tyr Arg	
1 5 10 15	
ttg cac ggc atc ttc ggc tcg cac gac acc acc tcg acc gcc ggt ggt	96
Leu His Gly Ile Phe Gly Ser His Asp Thr Thr Ser Thr Ala Gly Gly	
20 25 30	
gtc gcg aac gac atc aag ccg ttc aac ccc aaa cag gta acc ctc gag	144
Val Ala Asn Asp Ile Lys Pro Phe Asn Pro Lys Gln Val Thr Leu Glu	
35 40 45	
gtc ttt ggc gct ccc gga acc gtg gca acg atc aat tat ctg gac gtg	192
Val Phe Gly Ala Pro Gly Thr Val Ala Thr Ile Asn Tyr Leu Asp Val	
50 55 60	
gat gcc aca cct cgg caa gtc ctg gac acg acc ctg ccg tgg tca tac	240
Asp Ala Thr Pro Arg Gln Val Leu Asp Thr Thr Leu Pro Trp Ser Tyr	
65 70 75 80	
acg atc acg acg acc ctg ccc gcg gtc ttc gcc aat gtt gtc gcg caa	288
Thr Ile Thr Thr Thr Leu Pro Ala Val Phe Ala Asn Val Val Ala Gln	
85 90 95	
ggc gac agc aat tcc atc ggc tgc cgc atc acc gtc aac ggt gta gtc	336
Gly Asp Ser Asn Ser Ile Gly Cys Arg Ile Thr Val Asn Gly Val Val	
100 105 110	
aag gac gaa agg atc gtc aac gaa gtg cgc gcc tat acc ttc tgc ctc	384
Lys Asp Glu Arg Ile Val Asn Glu Val Arg Ala Tyr Thr Phe Cys Leu	
115 120 125	

gac aag tcc tca tga  
 Asp Lys Ser Ser  
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399

<210> 12  
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 <212> PRT  
 <213> Mycobacterium tuberculosis

<220>  
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 Leu His Gly Ile Phe Gly Ser His Asp Thr Thr Ser Thr Ala Gly Gly  
           20                  25                  30  
 Val Ala Asn Asp Ile Lys Pro Phe Asn Pro Lys Gln Val Thr Leu Glu  
           35                  40                  45  
 Val Phe Gly Ala Pro Gly Thr Val Ala Thr Ile Asn Tyr Leu Asp Val  
   50                  55                  60  
 Asp Ala Thr Pro Arg Gln Val Leu Asp Thr Thr Leu Pro Trp Ser Tyr  
   65                  70                  75                  80  
 Thr Ile Thr Thr Thr Leu Pro Ala Val Phe Ala Asn Val Val Ala Gln  
           85                  90                  95  
 Gly Asp Ser Asn Ser Ile Gly Cys Arg Ile Thr Val Asn Gly Val Val  
           100                  105                  110  
 Lys Asp Glu Arg Ile Val Asn Glu Val Arg Ala Tyr Thr Phe Cys Leu  
   115                  120                  125  
 Asp Lys Ser Ser  
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<210> 13  
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 <212> DNA  
 <213> Mycobacterium tuberculosis

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<210> 14  
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<400> 14  
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20

<210> 15  
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<400> 15  
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<210> 18  
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 <213> Mycobacterium tuberculosis

<400> 18  
 catagatccc ggacatggtg 20

<210> 19  
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 <212> DNA  
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<220>  
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 caacacgaac tcgccaaacg cttagaactt gggccgcagg cgaaatcggt ccagtcgccc 180  
 gagttcgccg ctcgcttgcc tgccgctcaa cacaggtagc gcctaccagc ctcgctgggtt 240  
 tccatggcgt gcccagtcg gaagctgctg ctgcttgact ccgcgcgctg ggcccagcgc 300  
 cgcgctgttg tacggcccaa acggcgtgtc ggtgtacagt cgcgcgctcg cggcttcagt 360  
 ccggcccccc gactccggca ggcccagcgg cgcccagcgc tagcccgaag ttcccccttg 420  
 taggggcggg ctgagtttcg atctgtttcg tgagcaggtg tttctgtggt caacttcctt 480

caacatgtac tcatgtatta ttgagaatag ctcggc	gtg tca tcc tct gat gac	534
	Val Ser Ser Ser Asp Asp	
	1 5	
gct att atc gcg ctg acc gcg tgt tat aaa gta atc atg tac att acc	582	
Ala Ile Ile Ala Leu Thr Ala Cys Tyr Lys Val Ile Met Tyr Ile Thr		
10 15 20		
cgg gta ccc aac cgg gga tcc ccg ccg gcg gtg ctg ttg cgg gaa agc	630	
Arg Val Pro Asn Arg Gly Ser Pro Pro Ala Val Leu Leu Arg Glu Ser		
25 30 35		
ttc cgc gaa aac ggc aag gtc aag acg cgt acc ctg gcc aac ctc tca	678	
Phe Arg Glu Asn Gly Lys Val Lys Thr Arg Thr Leu Ala Asn Leu Ser		
40 45 50		
cgc tgg ccc gag cac aag ctg gac aga ctg gac cgg gcg ctt aag ggc	726	
Arg Trp Pro Glu His Lys Leu Asp Arg Leu Asp Arg Ala Leu Lys Gly		
55 60 65 70		
ttg ccg ccc gcg gac tgg gat cta gcc gag gcc ttc gat atc acc cgc	774	
Leu Pro Pro Ala Asp Trp Asp Leu Ala Glu Ala Phe Asp Ile Thr Arg		
75 80 85		
agc ctg ccg cac ggg cat gtg gcc gcg gtg gcc ggc acc gcc gag aag	822	
Ser Leu Pro His Gly His Val Ala Ala Val Ala Gly Thr Ala Glu Lys		
90 95 100		
ctg ggc ata ccc gag ctg atc gac ccc acc ccg tcg cgg cgg cgc aac	870	
Leu Gly Ile Pro Glu Leu Ile Asp Pro Thr Pro Ser Arg Arg Arg Asn		
105 110 115		
ctg gtg ctg gcc atg ctg atc ggg cag atc atc gag ccc gga tcg aaa	918	
Leu Val Leu Ala Met Leu Ile Gly Gln Ile Ile Glu Pro Gly Ser Lys		
120 125 130		
ctg gcg atc gcg cgc ggg ctg cgc gcc cag acc gcc acc agc acg ctg	966	
Leu Ala Ile Ala Arg Gly Leu Arg Ala Gln Thr Ala Thr Ser Thr Leu		
135 140 145 150		
ggt gcg gtg ctg ggt gtc tcg ggc gcc gat gag gac gac ctg tat gac	1014	
Gly Ala Val Leu Gly Val Ser Gly Ala Asp Glu Asp Asp Leu Tyr Asp		
155 160 165		
gcg atg gac tgg gcg ctg gag cgc aaa gac ggc atc gaa aac gcc ttg	1062	
Ala Met Asp Trp Ala Leu Glu Arg Lys Asp Gly Ile Glu Asn Ala Leu		
170 175 180		
gcc gca cgg cat ctg acc aac ggc acc ctg gtg ctc tat gac gta tcc	1110	
Ala Ala Arg His Leu Thr Asn Gly Thr Leu Val Leu Tyr Asp Val Ser		
185 190 195		
tcg gcg gcg ttc gag ggc cac acc tgc ccg ctg gga gcg atc ggg cac	1158	
Ser Ala Ala Phe Glu Gly His Thr Cys Pro Leu Gly Ala Ile Gly His		
200 205 210		
gcc cgc gac ggg gtc aaa ggc cgg ctg cag atc gtc tac ggg ctg ctg	1206	
Ala Arg Asp Gly Val Lys Gly Arg Leu Gln Ile Val Tyr Gly Leu Leu		
215 220 225 230		

tgc tca ccc aag gga gcg ccg gtg gcc atc gag gtg ttc aag ggc aac	1254
Cys Ser Pro Lys Gly Ala Pro Val Ala Ile Glu Val Phe Lys Gly Asn	
235 240 245	
acc gcc gac ccg aaa act ctg aaa gct caa atc gac aag ctc aaa acc	1302
Thr Ala Asp Pro Lys Thr Leu Lys Ala Gln Ile Asp Lys Leu Lys Thr	
250 255 260	
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Arg Phe Gly Leu Thr Arg Ile Ala Leu Val Gly Asp Arg Gly Met Leu	
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Thr Ser Ala Arg Ile Arg Asp Glu Leu Arg Pro Ala His Leu Asp Trp	
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Ile Ser Ala Leu Arg Ala Pro Gln Ile Lys Ile Leu Leu Glu Asp Gly	
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Ala Leu Gln Leu Ser Leu Phe Asp Glu Gln Asn Leu Phe Glu Ile Thr	
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His Pro Asp Tyr Pro Gly Glu Arg Leu Val Cys Cys His Asn Pro Ala	
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Leu Ala Asp Glu Arg Ala Arg Lys Arg Ala Glu Leu Leu Ala Ala Thr	
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Glu Lys Glu Leu Gln Ala Ile Ala Glu Ala Thr Arg Arg Gln Arg Arg	
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Pro Leu Arg Gly Thr Asp Lys Ile Gly Leu Arg Val Gly Lys Val Arg	
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Asn Lys Phe Lys Met Ala Lys His Phe Asp Leu His Ile Thr Asp Glu	
395 400 405	
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Ala Phe Ser Phe Thr Arg Asn Gln Asn Ser Ile Ala Ala Glu Ala Ala	
410 415 420	
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Leu Asp Gly Ile Tyr Val Leu Arg Thr Ser Leu Pro Asp Asn Ala Leu	
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Gly Arg Asp Asp Val Val Gly Arg Tyr Lys Asp Leu Ala Asp Val Glu	
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Arg Phe Phe Arg Thr Leu Asn Ser Glu Leu Asp Val Arg Pro Ile Arg	
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cat ccg ctg gcc gac ccg gtc cgc gcc cac atg ttc ttg cac atg ctc	1974

His Arg Leu Ala Asp Arg Val Arg Ala His Met Phe Leu His Met Leu	
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Ser Tyr Tyr Ile Ser Trp His Met Lys Gln Ala Leu Ala Pro Ile Leu	
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ttc acc gac aac gac aaa ccc gcc gcc gcc gcc aaa cgc gcc gac ccc	2070
Phe Thr Asp Asn Asp Lys Pro Ala Ala Ala Lys Arg Ala Asp Pro	
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Val Ala Pro Ala Gln Arg Ser Asp Glu Ala Leu Asn Lys Ala Ala Arg	
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Lys Arg Thr Glu Asp Asn Gln Pro Val His Ser Phe Thr Ser Leu Leu	
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Thr Asp Leu Ala Thr Ile Cys Ala Asn Tyr Ile Gln Pro Thr Asp Asp	
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Leu Pro Ala Phe Thr Lys Thr Thr Thr Pro Thr Pro Thr Gln Arg Arg	
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Ala Phe Asp Leu Leu Ala Val Ser His Arg His Gly Leu Ala	
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35 40 45	
Thr Leu Ala Asn Leu Ser Arg Trp Pro Glu His Lys Leu Asp Arg Leu	
50 55 60	
Asp Arg Ala Leu Lys Gly Leu Pro Pro Ala Asp Trp Asp Leu Ala Glu	
65 70 75 80	
Ala Phe Asp Ile Thr Arg Ser Leu Pro His Gly His Val Ala Ala Val	
85 90 95	
Ala Gly Thr Ala Glu Lys Leu Gly Ile Pro Glu Leu Ile Asp Pro Thr	

100					105					110					
Pro	Ser	Arg	Arg	Arg	Asn	Leu	Val	Leu	Ala	Met	Leu	Ile	Gly	Gln	Ile
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Thr	Ala	Thr	Ser	Thr		Leu	Gly	Ala	Val	Leu	Gly	Val	Ser	Gly	Ala
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Glu	Asp	Asp	Leu	Tyr	Asp	Ala	Met	Asp	Trp	Ala	Leu	Glu	Arg	Lys	Asp
				165					170					175	
Gly	Ile	Glu	Asn	Ala	Leu	Ala	Ala	Arg	His	Leu	Thr	Asn	Gly	Thr	Leu
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Val	Leu	Tyr	Asp	Val	Ser	Ser	Ala	Ala	Phe	Glu	Gly	His	Thr	Cys	Pro
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Gly	Asp	Arg	Gly	Met	Leu	Thr	Ser	Ala	Arg	Ile	Arg	Asp	Glu	Leu	Arg
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Pro	Ala	His	Leu	Asp	Trp	Ile	Ser	Ala	Leu	Arg	Ala	Pro	Gln	Ile	Lys
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Asn	Leu	Phe	Glu	Ile	Thr	His	Pro	Asp	Tyr	Pro	Gly	Glu	Arg	Leu	Val
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Cys	Cys	His	Asn	Pro	Ala	Leu	Ala	Asp	Glu	Arg	Ala	Arg	Lys	Arg	Ala
			340					345					350		
Glu	Leu	Leu	Ala	Ala	Thr	Glu	Lys	Glu	Leu	Gln	Ala	Ile	Ala	Glu	Ala
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Thr	Arg	Arg	Gln	Arg	Arg	Pro	Leu	Arg	Gly	Thr	Asp	Lys	Ile	Gly	Leu
	370					375					380				
Arg	Val	Gly	Lys	Val	Arg	Asn	Lys	Phe	Lys	Met	Ala	Lys	His	Phe	Asp
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Leu	His	Ile	Thr	Asp	Glu	Ala	Phe	Ser	Phe	Thr	Arg	Asn	Gln	Asn	Ser
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Ile	Ala	Ala	Glu	Ala	Ala	Leu	Asp	Gly	Ile	Tyr	Val	Leu	Arg	Thr	Ser
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Leu Pro Asp Asn Ala Leu Gly Arg Asp Asp Val Val Gly Arg Tyr Lys  
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 Asp Leu Ala Asp Val Glu Arg Phe Phe Arg Thr Leu Asn Ser Glu Leu  
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 Asp Val Arg Pro Ile Arg His Arg Leu Ala Asp Arg Val Arg Ala His  
 465 470 475 480  
 Met Phe Leu His Met Leu Ser Tyr Tyr Ile Ser Trp His Met Lys Gln  
 485 490 495  
 Ala Leu Ala Pro Ile Leu Phe Thr Asp Asn Asp Lys Pro Ala Ala Ala  
 500 505 510  
 Ala Lys Arg Ala Asp Pro Val Ala Pro Ala Gln Arg Ser Asp Glu Ala  
 515 520 525  
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 Ser Phe Thr Ser Leu Leu Thr Asp Leu Ala Thr Ile Cys Ala Asn Tyr  
 545 550 555 560  
 Ile Gln Pro Thr Asp Asp Leu Pro Ala Phe Thr Lys Thr Thr Thr Pro  
 565 570 575  
 Thr Pro Thr Gln Arg Arg Ala Phe Asp Leu Leu Ala Val Ser His Arg  
 580 585 590  
 His Gly Leu Ala  
 595



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